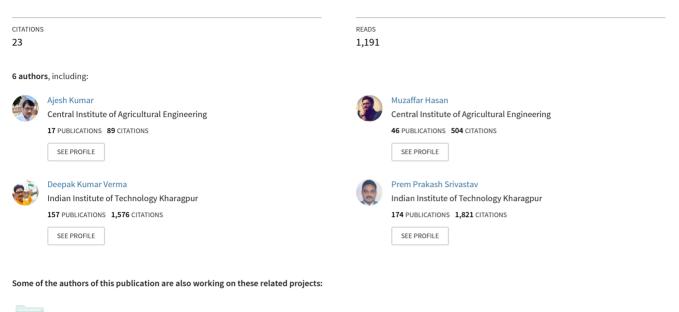
See discussions, stats, and author profiles for this publication at: https://www.researchgate.net/publication/360452134

# Trends in Edible Packaging Films and its Prospective Future in Food: A Review

Article · May 2022

DOI: 10.1016/j.afres.2022.100118



Project Prebioctic compounds View project
Project Science and Technology of Aroma, Flavor, and Fragrance in Rice View project

Contents lists available at ScienceDirect





Applied Food Research

journal homepage: www.elsevier.com/locate/afres

# Trends in Edible Packaging Films and its Prospective Future in Food: A Review



Ajesh Kumar V<sup>1,\*</sup>, Muzaffar Hasan<sup>2,\*\*</sup>, Shukadev Mangaraj<sup>1</sup>, Pravitha M<sup>2</sup>, Deepak Kumar Verma<sup>3</sup>, Prem Prakash Srivastav<sup>3</sup>

<sup>1</sup> Centre of Excellence for Soybean Processing and Utilization, ICAR-Central Institute of Agricultural Engineering, Nabibagh, Berasia Road, Bhopal 462 038, India <sup>2</sup> Agro Produce Processing Division, ICAR- Central Institute of Agricultural Engineering, Nabibagh, Berasia Road, Bhopal 462 038, India

<sup>3</sup> Agricultural and Food Engineering Department, Indian Institute of Technology Kharagpur, Kharagpur 721302, West Bengal, India

#### ARTICLE INFO

Keywords: Biopolymer Edible film Nanomaterials Composite film Safety and regulation

# ABSTRACT

Food packaging is an important area of food research due to its prime role in the protection and containment of foodstuffs. Traditionally petroleum-derived polymers fulfill the lion's share of packaging material requirements. However, present-day consumers are more concerned about the environmental impact and health hazards of these synthetic polymers. This necessitates the requirement of alternative packaging material with unique biodegradable and renewable characteristics. The edible film is considered a solution to replace these synthetic plastics with naturally available bio-macromolecules such as polysaccharides, proteins, and lipids. An enormous number of researches have been carried out across the world to explore its full potential. Their findings need to be consolidated for further development of this trending research area. Therefore, this article comprehensively reviews previous research progresses, such as different film formulations from various sources and their characteristics and product applications to guide the enthusiastic researchers. Finally, the last section of this article elaborates on safety and regulation aspects as well as recent trends and challenges to tackle all the obstacles in establishing a greener packaging option.

# 1. Introduction

The packaging is one of the most critical post-harvest operations for the preservation and shelf-life extension of fruits, vegetables, and processed foods. The major functions of food packaging include protection, communication, and convenience. Advancement in industrialization leads to the sharp growth in plastic use for food packaging. The production of plastic in the world has reached up to 380 million tonnes, and it has shown a steep increase in the past few decades, where 40% of the plastic produced is used in packaging applications (Groh et al., 2019). Although plastic is quite convenient as a packaging material, because of its low price, high mechanical strength, convenience in shape molding, heat sealability, and lighter in weight, enormous usage of plastic packaging material may lead to adverse effects on the environment (Cazón et al., 2017; Dehghani et al., 2018). For example, plastic waste virtually does not degrade, it will take hundreds of years for its disposal in a landfill, and the disposal of plastic through incineration can produce highly toxic gases (Otoni et al., 2017). Hence, plastic is considered the most significant menace in resolving earth pollution (Hasan et al., 2020).

In the past few decades, consumers are also aware of the impact of plastic on the environment. Therefore, the demands of alternate packaging materials which ensure an enhanced shelf-life with good quality and less impact on the environment are crucial in the food packaging industry. Edible packaging has been traditionally used to improve food appearance and preservation, and it captivated substantial attention in the last few decades due to the possibility of partial substitution of nonbiodegradable synthetic packaging materials (Hassan et al., 2018).

The primary role of edible film is controlling the moisture loss and reducing the adverse chemical reaction rates to enhance the quality and safety of a wide range of processed as well as fresh foods (Debeaufort, F. et al., 1998). In addition, the incorporation of various food additives such as antimicrobials, antioxidants, flavors, and colors into the edible film matrix further extends their applications (Tavassoli-Kafrani et al., 2016). However, the permeability and mechanical properties of the edible film are not on par with conventionally used synthetic plastic films

https://doi.org/10.1016/j.afres.2022.100118

Received 13 January 2022; Received in revised form 20 March 2022; Accepted 3 May 2022 Available online 7 May 2022

2772-5022/© 2022 The Author(s). Published by Elsevier B.V. This is an open access article under the CC BY license (http://creativecommons.org/licenses/by/4.0/)

<sup>\*</sup> Corresponding authors: Ajesh Kumar V, Centre of Excellence for Soybean Processing and Utilization, ICAR- Central Institute of Agricultural Engineering, Nabibagh, Berasia Road, Bhopal, India, Phone number: +91 8075913057.

<sup>\*\*</sup> Corresponding authors: Muzaffar Hasan, Agro Produce Processing Division, ICAR- Central Institute of Agricultural Engineering, Nabibagh, Berasia Road, Bhopal 462 038, India, Phone number: +91 8290767324.

E-mail addresses: ajeshmtr@gmail.com, ajesh.v@icar.gov.in (A.K. V), muzaffarhassan88@gmail.com (M. Hasan).



**Fig. 1.** Application of different types of edible film in various foods. a) Soybean aqueous extract-based nanocomposite edible film applied as cheese slice separator, b.) Guava (left) and beetroot (right) purees produced edible film, c.) Sodium alginate based edible film applied in meat slice, soluble coffee (d.), powder medicine (e.), cheese slice (V et al., 2021; Otoni et al., 2017; Gheorghita (Puscaselu) et al., 2020; Puscaselu et al., 2019).

(Murrieta-Martínez et al., 2018). Hence, the present research contributions were geared towards these property enhancements.

This article reviews the recent progress in edible packaging, emphasizing standalone edible films. Information including different types, formulation methods, application on food products, safety and regulations, and recent trends in the edible film is comprehensively reviewed. Finally, the challenges faced in commercial application of edible film in food packaging with possible solutions are also covered.

#### 2. Different materials used for edible film formation

Edible packaging materials are natural polymers obtained from polysaccharides, proteins (animal or vegetable), lipids, or combinations of these components (Khaoula et al., 2004; Galus & Kadzińska, 2015). According to Market Research Futures (MRFR), the edible packaging market (based on protein, lipids, polysaccharides, and other) will be worth USD 2.14 billion by 2030, with a compound annual growth rate (CAGR) of 6.79 percent (2022–2030), up from USD 783,32 million in 2021. North America will dominate the edible packaging market throughout the forecast period, followed by the United Kingdom, Japan, Indonesia, and Israel (Future, 2021). Several researchers formulated and characterized numerous edible films from different plant origin materials (Fig. 1). Researchers have continuously worked for the last three decades to develop edible films that can match the conventional plastic films to enable their commercial application. Varieties of edible packaging materials have been discussed in the following section.

# 2.1. Protein-based edible film

Edible packaging formed from proteins of plant origin includes corn zein, wheat gluten, soy protein, peanut protein, quinoa protein, sesame protein. In contrast, keratin, egg white protein, myofibrillar protein, collagen, gelatine, casein, and milk whey protein are film formers from animal sources (Mellinas et al., 2016). Among different edible film sources, protein-based material appears to be more attractive as they also provide nutritional value (Galus & Kadzińska, 2016). In addition, protein-based edible films have relatively higher mechanical and gas barrier properties with low moisture barrier properties. Protein-based films are better than lipid and polysaccharide films. They have excellent physical properties and gas-blocking effects because of their tightly packed and ordered hydrogen-bonded network structure (Kumari et al., 2017). Oxidation of lipid materials is the major cause of deterioration of quality and shelf life of high fat or fried foods. This can be controlled to a certain extent by using protein-based packaging, which inhibits oxygen permeation (Zhang, S & Zhao, 2017). The structure of protein also plays a crucial

role in oxygen permeability. It is reported that corn zein, wheat gluten, soy protein, and whey protein-based edible film have greater oxygen permeability than collagen-based films due to the globular structured proteins (Wittaya, 2012). Protein-based edible films can also be used for the individual packaging of small portions of food, particularly products for which individual packaging is not practically feasible, such as beans, nuts, and cashew nuts (Bourtoom, 2009). Both wet and dry methods can be used to create the protein-based film.

#### 2.1.1. Whey protein/milk protein

Whey protein or casein protein is preferred over the total milk protein for edible film formation as the latter results in crystallization due to the presence of lactose (Wagh et al., 2014). Edible films can be prepared from whey protein fraction, Whey Protein Isolate (WPI), and Whey Protein Concentrate (WPC) by adding different emulsifiers and plasticizers (Galus & Kadzińska, 2016; Soazo et al., 2016; Çakmak et al., 2020; Seydim et al., 2020). Whey protein film is characterized by its excellent oxygen, aroma, and oil barrier properties under low to medium relative humidity conditions. It also has the required mechanical properties for different applications like food coating, separating food layers, and pouch formation. In recent times, the addition of probiotics and prebiotics have been carried out to enhance the functional properties of the whey protein-based films (Fernandes et al., 2020; Zoghi et al., 2020).

#### 2.1.2. Wheat gluten protein

Wheat gluten, which contains more than 75% protein, is the protein part of the wheat flour after removing other starch granules by washing (Chavoshizadeh et al., 2020). It consists mostly of monomer gliadins and polymer glutenins in nearly equal amounts by weight. The cohesiveness and elasticity of gluten provide integrity and facilitate film formation (Fakhouri et al., 2017). Wheat gluten has very good oxygen and carbon dioxide barriers properties (Zubeldía et al., 2015). In addition, the ability to form cross-linking upon heating, visco-elasticity properties, low water solubility, low cost, and availability due to coproduct in the wheat starch industry make wheat gluten a favorite protein source for edible packaging (Ansorena et al., 2016). Although the wheat gluten-based films show brittleness and tendency to absorb water after being processed, the application of different methods such as adding plasticizers, incorporating additives possessing reactive groups (e.g. NH<sub>2</sub>, -COOH, -OH, and -SH), and blending with polymers (e.g; aliphatic polyester, poly (hydroxy ester ether), poly (lactic acid), polycaprolactone, poly (vinyl alcohol) and cassava starch) can minimize the impact (Hemsri et al., 2011). Moreover, the mechanical properties of gluten-based films are strongly affected by pH and gluten concentration, while water vapor permeability may be correlated with pH and

ethanol levels (Fakhouri et al., 2017). The physiochemical properties of the wheat gluten films can be improved by the incorporation of other different proteins, polysaccharides, and organic acids (Dong et al., 2022; He et al., 2020)

#### 2.1.3. Soy protein

Soy protein isolate (SPI) is one of the major sources of protein for edible packaging obtained from dehulled and defatted soybean (Cristine De Souza et al., 2020). Soy protein-based formation of the edible film occurred in two steps process- 1) disruption of soy protein complex structure through alkaline or heating treatment and cleavage of native disulfide bonds lead to exposure of sulfhydryl groups and hydrophobic groups; and 2) new disulfide bonds, hydrophilic and hydrophobic bonds formation. Utilizing native soy protein is challenging in applications like edible packaging due to its structural characteristics (Gao et al., 2015). However, modification of soy protein is possible with cross-linking of protein structure by different methods like denaturation, thermal treatment, and application of natural cross-linking agents (Friesen et al., 2015; Xia et al., 2015). Most commonly used protein cross-linkers are aldehydic compounds such as glutaraldehyde, formaldehyde, and glyoxal, phenolic, and epoxy compounds. The SPI-based edible films exhibit properties such as transparency, flexibility, low oxygen permeability, even comparable to low-density polyethylene film with abundant availability and low cost (Nandane & Jain, 2018). The gelling ability of the SPI makes it convenient in forming a suitable matrix for composite films with lipids as well as bioactive compounds such as antioxidants and antimicrobial agents (Carpiné et al., 2015; Cristine De Souza et al., 2020).

# 2.1.4. Sodium caseinate

Sodium caseinate (SC) is a water-soluble form of casein produced by adjusting acid-coagulated casein to pH 6.7 using sodium hydroxide (Belyamani et al., 2014; Yin et al., 2014). The randomly coiled structure of the SC enables good film formation (Lin, Wang and Weng, 2020). The earlier studies addressed that surface modification of SC film with zein coating has yielded film with better barrier properties (Yin et al., 2014). The structural inversion approach of zein coating on SC film resulted in surface irregularities with high irregular projections, which ultimately led to enhanced water and oxygen barrier properties. Whereas the direct coating only reduced the oxygen barrier property in which the zein nano-spheres were evenly distributed on the film surface. In recent studies, cross-linking with genipinin (Lin et al., 2020; Qiu et al., 2020), and incorporation of essential oils and antimicrobial agents (Alizadeh-Sani et al., 2020; Di Giuseppe et al., 2022) have proven to enhance the mechanical and antibacterial properties of the SC films.

#### 2.1.5. Corn zein

Corn zein is a major protein that can be utilized to prepare the edible film, edible coating, and pouches (Chen et al., 2014; Zhang & Zhao, 2017). Corn zein is extracted from corn gluten, a by-product of bioethanol production, which ensures plentiful availability (Escamilla-García et al., 2013). Zein is applied to other protein and polysaccharide-based films such as SPI and glucomannan (Wang Kai et al., 2017) to improve barrier properties and act as a finishing agent by imparting surface gloss (Cheng et al., 2015). The formation of zein coating on hydrophilic protein-based films via the specific protein-protein interactions has a promising potential to improve their barrier capability. Zein contains sharply defined hydrophobic and hydrophilic domains at its surface and is capable of self-assembly (Yin et al., 2014). The hydrophobic property of zein is contributed by the high proportion of non-polar amino acid residues, such as proline, leucine, and alanine. Thus, zein has been recommended as an edible film matrix material (Chen et al., 2014).

#### 2.1.6. Collagen and gelatin

Collagen, an animal-sourced protein used in edible packaging, is a hydrophilic protein rich in glycine, hydroxyproline, and proline; hence it swells in polar liquids with high solubility parameters (Coppola et al., 2020). Studies have reported that usage of collagen casing for meat products dates back to the 1920s (Janjarasskul & Krochta, 2010; Yang et al., 2016). Similar to collagen, gelatin is an animal protein obtained by controlled hydrolysis of the fibrous insoluble collagen present in the bones and skins generated as waste materials during animal slaughtering and processing (Lopez et al., 2017). The application of gelatin as an edible film has been extensively studied in several research studies (Bonilla & Sobral, 2016; Jridi et al., 2019). Gelatin is known for its application advantages such as good film-forming ability, good gas and oil resistance, nontoxicity, low price, and biodegradable properties. At the same time, its poor mechanical property, low thermal stability, weak water resistance, and rapid biodegradation property need to be improved (Ge et al., 2017). This can be tackled by forming a composite film using appropriate starch materials (Cheng et al., 2022).

#### 2.1.7. Other protein sources

For the sustainable use of protein sources, researchers started exploiting the use of protein from different sources to assess the edible film-forming abilities. These sources are chosen either as by-product utilization or for exploiting their unique properties and nutritional values. Protein from sources such as peanut protein and peanut protein isolate (Sun et al., 2013), lentil protein (Bamdad et al., 2006), a protein isolated from sesame (Sharma & Singh, 2016), myofibrillar proteins of fish muscle (Kaewprachu et al., 2016), pumpkin seed protein (Xu et al., 2019; Lalnunthari et al., 2020), egg white protein (Han et al., 2020; Huang X et al., 2020), and rice protein (Wang et al., 2020), etc. were also exploited for the formation of edible film. The major lacunae of these proteins are the inability to form a proper film-forming matrix. This can be overcome by cross-linking with transglutaminase and ultrasonication (Cruz-Diaz et al., 2019). These protein sources are used directly or in composite film formation.

#### 2.2. Polysaccharide-based edible film

Polysaccharides are the most abundant natural polymer, and recently, they have been widely used to prepare edible film or coatings materials (Imre et al., 2019). Polysaccharides *viz* cellulose, hemicellulose, starch, pectin, and derivatives of all these alginates, pullulan, chitin, and chitosan, are intensively used for edible film and coating materials preparation (Cazón et al., 2017). Polysaccharides-based edible films have a well-ordered hydrogen-bonded network, making them efficient oxygen blockers. However, polysaccharides-based films are less efficient in working as a moisture barrier due to their hydrophilic nature. Polysaccharide coatings are free from oil content, colorless in appearance, and used to extend the product's shelf life without creating any anaerobic condition (Mohamed et al., 2020). The polysaccharide-based film can be developed using both wet and dry methods. Commonly used polysaccharide materials for film formation include the following.

#### 2.2.1. Cellulose and its derivatives

Cellulose is the most abundant natural organic polymer, which can be applied for the preparation of the edible film. It is the primary structural component of the plant cell wall and is a linear homopolysaccharide comprised of  $\beta$ -1,4 glucose. Cellulose derivatives are derived from structural modifications like the addition of a small group (methyl, hydroxyl, and carboxyl) in cellulose (Fig. 2).

Principally four types of cellulose derivatives are used for edible coatings or films like Hydroxypropyl methylcellulose (HPMC; E464), Hydroxypropyl cellulose (HPC; E463), Methylcellulose (MC; E461), and Carboxymethylcellulose (CMC; E466) (Bourtoom, 2008). MC-based coatings create a barrier to in and out the movement of oil or lipids therefore used in confectionery foodstuffs. Similarly, HPMC-based film or coatings hinder the oil absorption consequently used for fried food products (Ngatirah et al., 2022)

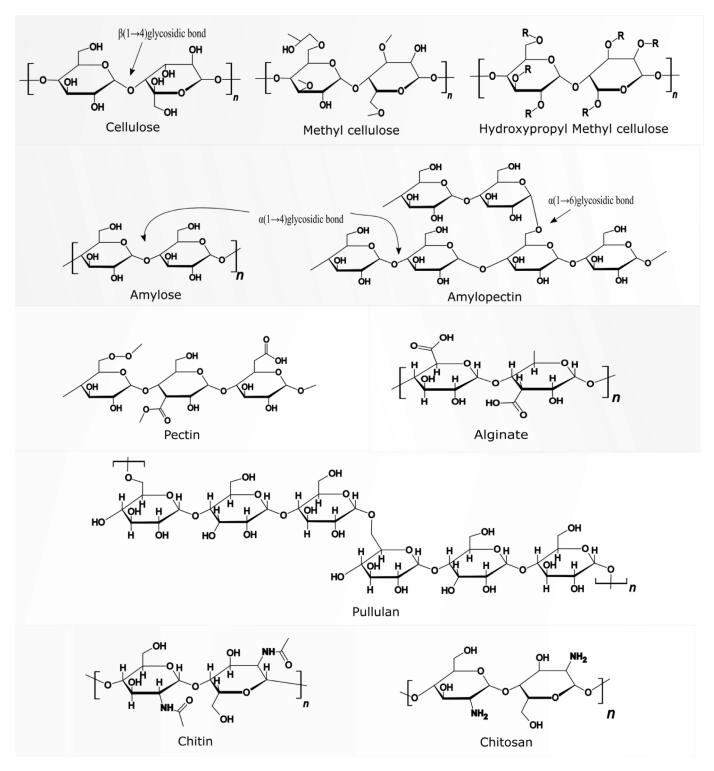


Fig. 2. Chemical Structure of different polysaccharides.

# 2.2.2. Starch and modified starch

Starch is a homo polysaccharide, comprised of amylose and amylopectin, used to develop biodegradable films because it can form a continuous matrix and is a renewable and abundant resource. Amylose is a linear polymer of  $(1\rightarrow 4)$  glucose, while amylopectin is a branched polymer of  $\alpha$  (1 $\rightarrow$ 4) glucose and  $\alpha$  (1 $\rightarrow$ 6) glucose (Fig. 2). Among amylose and amylopectin, amylose is generally used for film formation because of its high flexibility, low oxygen permeability, and water solubility (Cazón et al., 2017). However, the inherent hydrophilicity of amylose is marked by the solution of the solution of

lose makes it a poor barrier for water vapor. Therefore, improvising the property of the starch-based edible film, starch modification is needed (Askari et al., 2018).

# 2.2.3. Pectin

The polysaccharide pectin predominantly consists of galacturonic acid and its derivatives (Fig. 2). These polysaccharides are largely extracted from citrus peel and apple pomace (Morales-Contreras et al., 2020). The degree of esterification of pectin with methanol directly

affects the gelation and film-forming ability. Methoxy pectin (or esterified pectin) can be classified as low methoxy pectin (LMP) and high methoxy pectin (HMP) according to the degree of esterification (Espitia et al., 2016). Pectin is widely used in the edible film industry due to its biodegradability, biocompatibility, edibility, versatile chemical and physical properties such as selective gas permeability, and gelation (Chodijah et al., 2019)

#### 2.2.4. Alginate

Alginates have mannuronic acid and guluronic acid in structure (Fig. 2) and the composition of mannuronic acid and guluronic acid affects the physical property and molecular weight of alginates (Madsen et al., 2021). Alginate polysaccharides are mainly isolated from brown seaweeds. The colloidal nature of alginate, which includes thickening, stabilizing, film-forming, and suspending properties, makes it competent for edible film-forming material (Hassan et al., 2018). The presence of both the anionic sugar-acid enables it to bind divalent cations like Ca<sup>+2</sup>, Mg<sup>+2</sup>, Mn<sup>+2</sup>, and Fe<sup>+2</sup>. Therefore, incorporating divalent cations as a gelling agent causes the alginates-based edible film to be attributed to other physical properties like retaining moisture and color (Senturk Parreidt et al., 2018). The alginate-based edible film or coatings show lesser resistance to moisture or water because of the inherent hydrophilic nature of alginates (Dhanapal et al., 2012).

#### 2.2.5. Pullulan

The pullulan polysaccharide is secreted by the fungus *Auerobasidium pullulans* to resist desiccation and predation. The structure of pullulan mainly comprises maltotriose units (Fig. 2). Pullulan works as a thickener in edible film formation, and pullulans-based coatings are helpful to increase the shelf life of fruits (Diab et al., 2001). The edible coating property of pullulan can be enhanced by the use of glutathione (reducing agent) and chito oligosaccharide (antibacterial) in combination (Hassan et al., 2018). Like other polysaccharides, pullulan is hydrophilic in nature, which negatively affects its water barrier and mechanical properties. This limitation can be negotiated by adding lipids and fatty acids such as beeswax (BW), palmitic acid, and oleic acid (Omar-Aziz et al., 2021).

# 2.2.6. Chitin and chitosan

Chitin is the primary component of the cell wall of fungi and invertebrates. Deacetylation of chitin in alkali solution, convert it into chitosan. The repetitive unit in chitin polysaccharide is *N*-acetylglucosamine, a derivative of glucose monosaccharide (Fig. 2). Chitosan-based edible films have barrier properties for  $O_2$  and  $CO_2$  as well as possess inherent antimicrobial properties. The physicochemical properties of chitosanbased edible film or coatings are varied with the degree of deacetylation of chitin (Kumar et al., 2020). Chitin is characterized by its excellent biodegradability, biocompatibility, antibacterial activity, and low immunogenicity (Li et al., 2019). Chitosan products are highly viscous, resembling natural gums with antimicrobial properties due to active amino groups (Nguyen et al., 2020), and they can form transparent films to enhance the quality and extend the storage life of food products (Ribeiro et al., 2020).

#### 2.3. Lipids-based edible film

Lipids are naturally originated compounds from plants, animals, and insects. The diversity of the lipid functional groups is made up of mono-, di-, tri-glycerides, phospholipids, phosphatides, terpenes, cerebrosides, fatty acids, and fatty alcohol (Mohamed et al., 2020). Unlike protein and polysaccharides, lipids alone cannot form an edible film. Though they are capable of forming an edible coating, the lack of a large number of repeating units connected by covalent bonds prevents the formation of a stand-alone film. Therefore, different plant and animal-based lipids (oils and fats) are incorporated in film-forming solution (FFS) to the emulsion-based edible film to impart more hydrophobic properties due to their low polarity (Janjarasskul & Krochta, 2010; Galus & Kadzińska, 2015). Oils and fats are chemically similar mixtures where main components are triglycerides but differ in origin and physical appearance. Oils come from plants and liquids in nature, whereas fats originate from animals and are solid in appearance at room temperature. Different vegetable oils (sunflower oil, olive oil, rapeseed oil, etc.), plant-based waxes (candelilla, carnauba, and sugar cane waxes), animalbased waxes (beeswax, lanolin, and wool grease), and synthetic waxes like paraffin wax and petroleum wax were added to form FFS (Rhim & Shellhammer, 2005). Waxes are made up of alcohol and/or esters of a long-chain acid; therefore, waxes have a larger molecular weight with potent hydrophobicity. Vegetable oil is a raw, oily material produced from nuts, seeds, or newly cut flowers after being pressed cold. On the other hand, essential oil is a highly aromatic compound produced through steam distillation from various parts of a plant (leaves, roots, fruit, wood, and flower). Essential oils are highly rich in hydrophobic, aromatic, and volatile compounds like terpenes and terpenoids. Furthermore, essential oils have potent antimicrobial properties.

Some lipid materials like virgin coconut oil added to the FFS have reduced the water vapor permeability (WVP) of the edible film prepared from SPI (Carpiné et al., 2015; Fangfang et al., 2020). To give antioxidant and antimicrobial properties, essential oils from many aromatic spices of clove, rosemary, cinnamon, lemon, thyme, garlic, oregano were added into FFS during the emulsification process. Encapsulating essential oils in the edible film also gives them stability against their volatile nature (Alexandre et al., 2016; Perdones et al., 2016; Hashemi & Mousavi Khaneghah, 2017). The main disadvantage of lipid film material is its fragile nature. It also makes the film waxy and greasy in texture and taste, which is not desirable for packaging material on many occasions. To have the desired properties for the film, the compatibility of the lipid phase with the polymer matrix is essential. The use of different essential oils in edible film formulation for improving the WVP as well as the functional properties such as antibacterial and antioxidant properties are listed in Table 1. The lipid-based self-supporting edible film is generally prepared with an FFS containing any of the high molecular weight polymers (protein or polysaccharide) using the solvent casting method (Rhim & Shellhammer, 2005).

#### 2.4. Composite edible film

Composite films are multi-component systems in which different hydrophobic, as well as hydrophilic compounds are blended to achieve better functional properties. Many times, single functional compounds forming the polymer matrix, which is capable of forming a structural matrix with sufficient cohesiveness, may not be sufficient to provide all of the required properties, such as mechanical, barrier, and so on (V. et al., 2022; Dhumal & Sarkar, 2018). Polar bio-polymeric edible films like polysaccharides and protein generally show good gas barrier properties and reasonably good mechanical properties at low relative humidity. Nevertheless, they show poor water barrier properties due to their hydrophilic nature at high humidity. In contrast, hydrophobic lipids are reasonably efficient against moisture migration, but due to their non-polymeric nature, they show poor mechanical properties and are inferior to those of hydrocolloid films (Janjarasskul & Krochta, 2010). So, mixing this hydrophobic lipid with hydrophilic polysaccharides or protein can yield films with better properties than those formed from individual compounds. For example, Omar-Aziz et al. (2021) experimented with developing film by combining beeswax and pullulan. They have found a significant improvement in WVP and TS in the composite film than those films formed from pullulan alone.

Composite films are prepared either in layer form or in the emulsion of film-forming materials. Layered composite films are classified into binary or ternary based on the number of polymers used. Several combinations of carbohydrate-protein (Wang Kun et al., 2017; Tavares et al.,

#### Table 1

Use of different essential oils in edible film formulation.

Matrix Polymer	Essential Oil Used	Targeted Product	Film Formation Method	Observations and Remarks	Reference
Carboxymethyl chitosan: Pullulan	Galangal essential oil (GEO)	Mango	Casting	Developed film exhibited excellent thermal stability, biodegradability and mechanical properties and was able to provide good preservation effect on mango.	(Zhou et al., 2021)
Gelatin: Green tea extract	Lemon essential oil (LEO)		Casting	Incorporation of green tea extract and LEO helps to achieve good WVP for the developed film.	(Nunes et al., 2020)
Gelatin–chitosan blend	Ferulago angulate essential oil (FAEO)	Turkey meat	Casting	FAEO incorporated in gelatin-chitosan blend film improved the water solubility and WVP. Increased anti-microbial property of the film helped in enhancing the shelf life of turkey meat.	(Naseri et al., 2020)
Millet starch	Clove essential oil		Casting	Inclusion of clove oil enhanced the anti-oxidant activity and antimicrobial properties of the film.	(Al-Hashimi et al., 2020)
SPI-gum acacia conjugates	Oregano essential oil (OG-EO), lemon essential oil (LM-EO), fruit of Amomum tsaoko Crevost et Lemaire (ACL-EO) and/or grapefruit essential oil (GF-EO)		Casting	GF-EO contained film exhibited better WVP, mechanical properties and glass transition temperature than other EO containing films. However, radical scavenging activity and antimicrobial activity was superior for LM-EO incorporated films.	(Xue et al., 2019)
Basil seed gum	Oregano essential oil		Casting	The resulting film showed a significant reduction in WVP with antimicrobial and antioxidant activity.	(Hashemi & Mousavi Khaneghah, 201
SPI:Acetem: Tween 60	Carvacrol and cinnamaldehyde		Casting	The addition of emulsions significantly reduced the tensile strength of the films and improved their EAB. An only slight improvement is reported with the addition of essential oils	(Otoni et al., 2016)
Gelatin: MMT	Ginger essential oil (GEO)		Casting	Synergetic effect of GEO with MMT significantly improved the mechanical properties like EAB, puncture force and puncture deformation.	(Alexandre et al., 2016)
Zein	Zataria multiflora Boiss. essential oil (ZEO)	Minced meat	Casting	Addition of ZEO along with monolaurin significantly improved the antioxidant activity and antimicrobial properties against <i>L. monocytogenes and E. Coli</i>	(Moradi et al., 2016)
Chitosan	Cinnamon and Ginger essential oil	Pork	Casting	Cinnamon and Ginger essential oil has distinctly increased the thickness and opacity of the chitosan films. The WVP of films remained unaffected. Incorporating 1% EOs yielded the highest antimicrobial and antioxidant activities chitosan films	(Wang et al., 2017)
WPI	Almond and walnut oils		Casting	Addition of oils increased the opacity of the film whereas swelling, water vapor permeability, and surface hydrophilicity were reduced.	(Galus & Kadzińska, 2016)
Chitosan:MMT	Rosemary essential oil and ginger essential oil	fresh poultry meat	Casting	Incorporation EOs improved only the barrier to oxidation but not the antimicrobial properties. Overall performance of EOs in Chitosan/MMT film is not significant	(Pires et al., 2018)

Abbreviations: EAB: Elongation at break, EO: Essential oil, MMT: Montmorillonite WVP: Water vapor permeability, SPI: Soy protein isolate, WPI: Whey protein isolate.

2021), protein-protein (Dong et al., 2022; Tsai & Weng, 2019), and carbohydrate–carbohydrate (Cheng et al., 2015; Fan, Yang, Duan, & Li, 2021) is possible in the case of binary film. A myriad of literature is available in the case of binary film, but limited numbers of works are reported in case of ternary composite edible films (Dhumal & Sarkar, 2018). Composite films formed by emulsifying the constituents give the better film than layered ones, as the layered film may tend to delaminate over time, and it also requires a greater number of casting and drying processes (Galus & Kadzińska, 2015). Composite film prepared by emulsifying the lipid phase into the hydrocolloid-based structural matrix provides better functionality and barrier properties (Ochoa et al., 2017). The proportion of various polysaccharides in the composite film can also influence the different physical and optical properties to an ex-

tent (Saberi et al., 2016). Several studies have been published on the composite film made by combining polysaccharides and lipids, with the goal of improving its water barrier properties. The size of the lipid particle on the composite film had a significant effect on WVP properties and mechanical properties due to its higher surface area (Otoni et al., 2016). It is also found that composite films using minerals and protein have shown a significant improvement in their mechanical properties (Wang et al., 2015).

Bi-layer films, predominant in composite edible films, can be prepared by casting one layer over the other, but this multi-step process involves the risk of layer delamination. A single-step process in which one compound is dispersed with other FFS and then cast to form the film can solve this problem (Valencia-Sullca et al., 2018; Zuo et al., 2019). The incorporation of essential oil in nano-emulsion form is the recent trend followed by the researcher for better efficacy (Shen et al., 2021). Details of some composite edible films are given in Table 2.

### 2.5. Nanoparticle based edible films

In the last decade, nanotechnology has been used as an innovative approach to obtaining nano-scaled organic and inorganic compounds with unique properties due to their size (Espitia & Otoni, 2018). By definition, nanomaterials have at least one of its dimensional particle sizes of about 1-100 nm. The application of nanomaterials in the food packaging sector is an emerging area. Incorporating nanomaterials into matrix polymers proved to be a promising strategy for improving their physical and mechanical properties, which conventional components cannot achieve (Bizymis & Tzia, 2021). Moreover, it can be used for the synthesis of efficient active packaging materials by the method of nanoencapsulation of bioactive natural materials. The nanoencapsulation technique improves the stability and solubility of bioactive compounds, thus leading to the formation of an active film with better performance than conventional ones (Pal et al., 2017). Various researchers have reported several combinations of the bio-based matrix polymer and nanoparticles. The application of nano clay is widely exploited due to its ability to improve the barrier and mechanical properties by their high aspect ratio and surface to volume ratio (Shekarabi et al., 2014). Çağrı Mehmetoğlu et al. (2021) demonstrated the effect of silver nanoparticles in whey protein-based films. Their findings show that adding silver nanoparticles to a film increases its tensile strength by 84% and its barrier properties by 67% over a control film. Similarly, other nanoparticles like zinc oxide, titanium oxide, nano cellulose, etc., have been widely used in food packaging (Dash et al., 2019; Malik & Mitra, 2021; Yekta et al., 2020).

However, whether these nanomaterials are safe or not is still a controversial question for the scientific community. Nanomaterials can have various toxic effects depending on their chemical composition, particle size distribution, particle shape, and surface condition. The potential to cause oxidative stress and, in some cases, inflammatory responses or genotoxic effects are the most common effects observed in experimental studies. The intensity of this harmful effect further depends on the nanomaterial dose in that particular FFS (Malakar et al., 2021). Based on the size, their ability to penetrate the human cells also varies. For example, 100 nm particles can easily penetrate cells, 40 nm can enter nuclei, and below 35 nm can cross the blood-brain barrier. Moreover, smaller sized particles will have more catalytic ability, and their reactive oxygen species producing potential, adsorption rate, and binding capability may be comparatively higher than bigger-sized particles (Bumbudsanpharoke et al., 2015; Vlachogianni & Valavanidis, 2014). Reliable data on nanoparticles' safety and toxicological effects is still not available in the public domain. Hence, the effect of these nanoparticles on human health and environmental microbiota needs to be explored in detail to rule out any adverse effect.

# 3. Methods for edible film formulation

The edible film can be prepared mainly by the wet process and dry process (Fig. 3). In the wet process, biopolymers are solubilized or dispersed in an aqueous solution, water-based or alcohol-based, to form FFS followed by drying of the solvent. In the dry process, biopolymers were converted into the film by utilizing the thermoplastic behavior exhibited by some proteins and polysaccharides at low moisture levels (Cao et al., 2007; Nussinovitch, 2013).

The wet process, also known as solvent casting, is the most predominant technique used in edible film formulation (Fig. 4 A). The solvent casting process involves the following steps described by Rodríguez et al. (2020), they are as follows: 1) solubilizing the base biopolymer into a suitable solvent such as water or ethanol to form a FFS, 2) casting the FFS into suitable moulds or Teflon coated plates, 3) drying the casted film formulation solution, 4) peeling/removing the film and storing at suitable RH and temperature.

During the formulation of FFS, all the components are mixed in to homogenize solution with the help of low-speed stirrings, ultrasonication sometimes at a higher temperature suited for the solubilization of the components into the solvents (Abral et al., 2019). The FFS should be free from air bubbles to avoid the entrapment in the film matrix that can affect the structural integrity of the film. Air bubbles from low viscous FFS are normally removed by vacuum degassing (Ghasemlou et al., 2011; Kim & Min, 2012; Jouki et al., 2013). In the casting step, the amount of the solution is controlled for adjusting the film thickness. For example, V et al. (2022) developed an edible film from soybean aqueous extract by following steps like initial mixing of beeswax, clove essential oil, and span-20 using a magnetic stirrer and ultrasonication followed by solution casting on Teflon sheet and drying at ambient temperature. As of now, the majority of the works are limited to bench casting, which is a batch process. Some limited research on continuous casting is also reported. In the drying process, the solvent is evaporated to form an edible film. The drying is usually carried out at ambient air condition or at low temperature (below 60°C) in a hot air dryer. The drying condition plays a major role in determining the properties of the film (Bagheri et al., 2019). Although the drying time is considerably reduced by alternative drying methods such as microwave and IR drying, it had a significant negative effect on the quality and mechanical properties without affecting WVP of the film (Kaya & Kaya, 2000; Srinivasa et al., 2004; Cárdenas et al., 2008; Tapia-Blácido et al., 2013).

The dry process is mainly classified into extrusion, compression molding, and injection molding. The extrusion method is widely used for commercial synthetic plastic film formulation (Fig. 4 B), in which the film-forming matrix is subjected to structural changes by the effect of high temperature, pressure, and low moisture content (Hernandez-Izquierdo & Krochta, 2008; Dang & Yoksan, 2015). In this method, the edible bioplastic materials are first converted into pellets and extruded with suitable plasticizers (Huntrakul et al., 2020; Vedove et al., 2021). The film formulation through extrusion occurs in three steps; i) feeding the FFS to the extruder, ii) mixing of the FFS in the kneading zone of the extruder, iii) heating the FFS and either passing through the slit die followed by calendaring (slit-die extrusion) or blowing through circle die (blown-film extrusion).

The process variables in the extrusion of films, such as screw speed, temperature, feeding rate, and moisture content, have shown great influence on the properties of the film (Jebalia et al., 2019; Ochoa-Yepes et al., 2019). Extrusion of the film is considered as the most suitable for the large-scale commercial production of edible film with low energy consumption and short processing time. However, the high temperature generated during the extrusion causes undesirable changes in the biopolymer, such as nutritional and sensory losses in edible film, and application is limited to a certain polymer that is tolerant to high temperature with low moisture content FFS (Otoni et al., 2017; Suhag et al., 2020).

Compression molding is considered a sustainable process compared to the traditional solvent casting method due to its rapid formation and less energy requirement (Uranga et al., 2018). In compression molding, film-forming materials are subjected to high pressure and temperature in the mold until solidification (Lisitsyn et al., 2021). Processing parameters like temperature, pressure and time are critical in deciding the film properties. The compression method is frequently used with the extrusion method, in which the former is used for preparing the film-forming material prior to the thermoforming process in the latter. Ceballos et al., (2020) developed an edible film from cassava starch and yerba extract. The ingredients were extruded into thread form using a twin-screw extruder, followed by compression molding to yield the film. A compression-molded film can have higher thickness and more flexibility than solvent cast film (Krishna et al., 2012).

The injection molding method is popular for the industrial production of plastics. It is suitable for the mass production of edible films.

# Table 2

Composite edible film using different polymer matrices.

Matrix Polymer	Plasticizer	Method of Preparation	Observations and Remarks	Reference
Ipomoea batatas: k-carrageenan	Glycerol	Casting	Composite blend of <i>Ipomoea batatas</i> and $\kappa$ -carrageenan yield film with good mechanical and optical properties.	(Bharti et al., 2020)
Chitosan: Nano-silicon aerogel: Okra powder	Glycerol	Casting	Combination of chitosan, Nano-silicon aerogel with okra powder improved	(Lin et al., 2020)
			mechanical, barrier, optical and anti-microbial properties with excellent	
			surface characteristics.	
Pearl millet starch: Carrageenan gum	Glycerol	Casting	Starch and carrageenan concentration has positive influence on the Tensile strength	(Sandhu et al., 2020)
			and barrier properties of the film. It enhanced optical, WVP and mechanical	
Chitosan: WPI	Glycerol	Casting	property values of the film. Composite film resulted in high tensile	(Tavares et al., 2021)
Shitosan. wrri	Giyceloi	Castilig	strength, lower deformation, flexibility,	(1avales et al., 2021)
			malleability and good WVP than the films formed individually.	
PG: Modified starch		Casting	Incorporation of PG reduced WVP, WS,	(Askari et al., 2018)
		~	MC, and TS of the composite film with an increase in its % EB especially when gum	
			percentage greater than 50. Result of	
			Morphological analysis shows that developed film is having good	
	et i		homogeneity with smooth structure.	
SPI: VCO:SL	Glycerol	Casting	Amalgamation of SPI, VCO and SL resulted in the formation of film with	(Carpiné et al., 2015)
			increased EAB and lower MC than the	
			film obtained from SPI alone. However, this blending was not influenced the WVP value of the film.	
Gelatin: DXG: NH <sub>2</sub> -MMT	-	Casting	Cross-linking effect of DXG and NH2-MMT	(Ge et al., 2017)
delatin. DAd. 1112-10101			nanofiller resulted in the enhancement of	(,, )
			water resistance, UV barrier property, and mechanical properties of the gelatin-based	
			composite film. Enhanced hydrophobicity	
			and compact structure resulting from the	
			cross-linking slowed down the fungal degradation of the film.	
Gelatin: GLU	Glycerol,	Casting	Gelatin in the composite film improved its	(Fakhouri et al., 2017)
	Sorbitol	custing	WVP and gluten enhanced its flexibility. EAB was greatly influenced by glycerol	
			concentration. However, presence of	
	Classes 1	Centing	Sorbitol did not alter the EAB.	(Table of the YY of the
Fish gelatin: Chitosan	Glycerol	Casting	Addition of glycerol caused significant increase in the TS and elastic modulus,	(Fakhreddin Hosseini et al., 2013)
			leading to stronger films as compared with gelatin film, but significantly	et al., 2013)
			decreased the EAB. Chitosan drastically	
			reduced the WVP and solubility of gelatin films.	
Gelatin: Starch: $\epsilon$ -PL	Glycerol	Extrusion blowing	Starch/gelatin mix is a suitable substrate	(Cheng et al., 2022)
			for making $\epsilon$ -PL loaded antimicrobial edible packaging.	
Cassava starch: Pea	Glycerol	Extrusion blowing	Incorporation of pea protein isolate at	(Huntrakul et al., 2020)
Cassava starch: Pea protein	01,00101	Life doton blowing	20% in cassava starch increased the strength of the composite film.	(Tunitunui et un, 2020)
Casein: wax powder:	Glycerol	Extrusion blowing	Extrusion was used to create a new	(Chevalier et al., 2018)
ootassium sorbate		0	composite edible film based on casein and	
Cassava starch: YME	Glycerol	Compression molding	several edible waxes. Tensile toughness of the cassava starch	(Ceballos et al., 2020)
Jassava Statuti. TIME	Giyter01	Compression molding	YME composite film highest was obtained for 10% YME	(Cenanos et al., 2020)
GLU: MMT	Glycerol	Injection molding	MMT showed and lubricating effect and	(Cho et al., 2011)
	,		facilitated the injection molding of GLU	(, =011)

Abbreviations: DXG: Dialdehyde xanthan gum EAB: Elongation at break, GLU: Wheat gluten, MC: Moisture content, NH<sub>2</sub>-MMT: Amino-functionalized montmorillonite, PG: Psyllium Gum, SL: Soy lecithin, TS: Tensile strength, VCO: Virgin coconut oil, WPI: Whey protein isolate, WS: Water solubility, WVP: Water vapor permeability, YME: yerba mate extract, ε-PL: ε-polylysine hydrochloride.

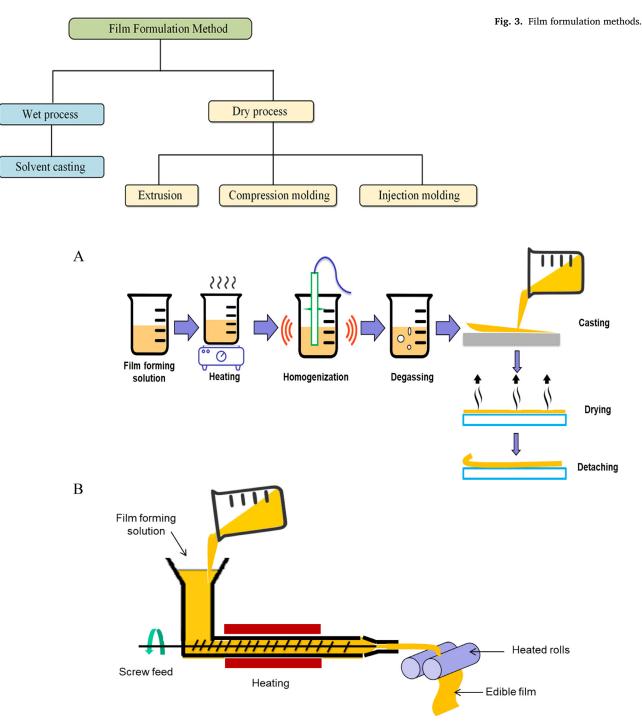


Fig. 4. Schematic representation of the edible film formulation: (A) Casting method and (B) extrusion method.

This method is used in combination with the extrusion method for obtaining the final film (Mellinas et al., 2016). However, limited research was reported in edible film injection molding (Mellinas et al., 2016). Cho et al. (2011) demonstrated the successful development of wheat gluten-based nanocomposite film using injection molding. The ingredients were pelletized with the help of a compression mold prior to injection molding using a three-phase screw injection-molding machine. Among the processing parameters, pre-injection temperature, molding temperature, and injection pressure are most critical for injection molding (Perez et al., 2016).

# 4. Application of edible film for packaging of foods

Every year the demand for the packaging material increases by 8% to meet the total requirement (Rodrigues et al., 2016; Tavassoli-Kafrani et al., 2016). This increases, and consumers' concern over green packaging triggers the spike in the utilization of bio-based food packaging materials for shelf-life extension of different food products (Falguera et al., 2011). Some of the successful applications of edible film explored to date are elaborated in the proceeding sections, and Table 3 summarizes the different applications of edible packaging on various food types.

#### Table 3

Application of edible film/coatings for essential functions in fruits, vegetables, meat and seafood.

(A) Edible Packaging of Frui		Food Commodities	Significant Function	References
.,	ts			
Candielila wax	Mineral oil	Guava fruits	Color retention, weight loss	(Tomás et al., 2005)
Cabdielila wax	Jojuba oil + ellagic acid	Golden delicious apple	Weight loss and Sensory qualities	Ochoa et al., 2011
Candelilla wax	Guar gum +glycerol	Strawberry	Anti-fungal, increase shelf life	(Oregel-Zamudio et al., 2017)
Carnauba wax		Walnuts, pine nuts	Overall appearance by reduction in rancidity, taste	(Mehyar et al., 2012)
Polysaccharide + car-		Mango	During the storage of mango ripening	(Baldwin et al., 1999)
nauba		0	improve the permeability	
wax			1	
Carrageenan + whey	CMC salt+	Apple	Browning on minimally processed	(Lee et al., 2003)
• •	PEG + CaCl2 + glycerol + ox-		apple slices	(,,,,
protein			apple silces	
	alic acid			
	additives			
Pectin + LDH-salicylate	Glycerol	Apricot	Morphological, thermal and barrier properties	(Gorrasi & Bugatti, 2016)
Whey protein + pectin	Sorbitol + gallic	Fresh apples, carrots,	Antioxidant, phenolic content, weight	(Rossi Marquez et al., 2017)
·····)	acid + transglutaminase	potatoes	loss, texture and sensory attribute	(,,, )
	-	-	-	
HPMC + lipid	potassium sorbate + sodium	Oranges	Improved antifungal property during	(Valencia-Chamorro et al., 2009
	benzoate + sodium		storage	
	propionate + stearic			
	acid + glycerol additives			
Cabdialila mar		Colden delietere en le	Compound qualities and the loss	(Herror et al. 2010)
Cabdielila wax	Jojuba oil + ellagic acid	Golden delicious apple	Sensory qualities, wight loss	(Hassan et al., 2018)
Candelilla wax	Guar gum +glycerol	Strawberry	Anti-fungal, increase shelf life	(Hassan et al., 2018)
Carnauba wax		Walnuts, pine nuts	Hydrolytic and oxidative rancidity,	(Bhartiya et al., 2020)
		, price nato		= 00 un, 2020)
(D) D 1111 D 1 1 1 1 1	. 11		taste, overall appearance	
(B) Edible Packaging of Vege	tables			
Chitosan + gelatin		Red bell peppers	firmness, $CO_2$ , weight loss, and ethanol concentration	(Poverenov et al., 2014)
Calcium caseinate	Acetylated	Zucchini (summer	reduce water	(Hassan et al., 2018)
			leuuce water	(Hassall et al., 2016)
coatings	monoglycerides + fatty acid	squash)		
	esters + sodium salt of			
	carboxymethyl cellulose			
Candelilla wax	carbonymentyr centatore	Prussels enrout	Paduage weight loss Procernation of	(Vouvologyik 2011)
Landenna wax		Brussels sprout	Reduces weight loss, Preservation of	(Kowalczyk, 2011)
			vitamin C and polyphenols, mini-	
			mal softening, increased overall ap-	
			pearance during	
			storage.	
HPMC + Beeswax	Oleic acid + glycerol	Cherry tomato	Weight loss, peel color, fruit firmness,	(Fagundes et al., 2015)
			respiration rate, sensory qualities	
			enhanced.	
k-carrageenan or Tapioca	Potassium sorbate + ascorbic	Fortified numplin	Color and antimicrobial activities	(Genevois et al., 2016)
		Fortified pumpkin		(Genevois et al., 2010)
			Fresh-cut	
starch coatings	acid + citric acid + glycerol			
starch coatings				
starch coatings (C) Edible Packaging for Sea	Foods and Meat Products	Salmon	Reduce microbial growth Reduce	(Alves et al., 2017)
starch coatings (C) Edible Packaging for Sea	Foods and Meat Products 0.8% glycerol+ Tween	Salmon	Reduce microbial growth Reduce	(Alves et al., 2017)
starch coatings (C) Edible Packaging for Sea	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed	Salmon	Reduce microbial growth Reduce lipid oxidation	(Alves et al., 2017)
starch coatings (C) Edible Packaging for Sea	Foods and Meat Products 0.8% glycerol+ Tween	Salmon	-	(Alves et al., 2017)
starch coatings ( <b>C</b> ) <i>Edible Packaging for Sea</i> 1.25 % chitosan	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol	Salmon Salmon	lipid oxidation	(Alves et al., 2017) (Castro et al., 2019)
starch coatings ( <b>C</b> ) <i>Edible Packaging for Sea</i> 1.25 % chitosan Whey protein	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed		-	
starch coatings (C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract	Salmon	lipid oxidation Retarded lipid oxidation	(Castro et al., 2019)
starch coatings (C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol		lipid oxidation Retarded lipid oxidation Microbial growth inhibited by olive	
(C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate Gelatin	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract	Salmon	lipid oxidation Retarded lipid oxidation	(Castro et al., 2019)
starch coatings (C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate Gelatin	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract	Salmon	lipid oxidation Retarded lipid oxidation Microbial growth inhibited by olive	(Castro et al., 2019)
starch coatings (C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract Olive leaf extract+glycerol	Salmon Cold smoked Salmon Cold smoked wild pacific	lipid oxidation Retarded lipid oxidation Microbial growth inhibited by olive leaf extracts	(Castro et al., 2019) (Albertos et al., 2017)
starch coatings (C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate Gelatin Chitosan (94.7% DD)	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract Olive leaf extract+glycerol Bacteriocin (divergicin M35)	Salmon Cold smoked Salmon Cold smoked wild pacific sockeye salmon	lipid oxidation Retarded lipid oxidation Microbial growth inhibited by olive leaf extracts Effective against L. monocytogenes	(Castro et al., 2019) (Albertos et al., 2017) (Benabbou et al., 2018)
starch coatings (C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate Gelatin Chitosan (94.7% DD) 2 % chitosan (75-85 %	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract Olive leaf extract+glycerol Bacteriocin (divergicin M35) Glycerol + tween 80+ grape	Salmon Cold smoked Salmon Cold smoked wild pacific	lipid oxidation Retarded lipid oxidation Microbial growth inhibited by olive leaf extracts Effective against L. monocytogenes Improved shelf life by Deferring	(Castro et al., 2019) (Albertos et al., 2017)
starch coatings (C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate Gelatin Chitosan (94.7% DD) 2 % chitosan (75-85 %	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract Olive leaf extract+glycerol Bacteriocin (divergicin M35)	Salmon Cold smoked Salmon Cold smoked wild pacific sockeye salmon	lipid oxidation Retarded lipid oxidation Microbial growth inhibited by olive leaf extracts Effective against L. monocytogenes	(Castro et al., 2019) (Albertos et al., 2017) (Benabbou et al., 2018)
starch coatings (C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate Gelatin	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract Olive leaf extract+glycerol Bacteriocin (divergicin M35) Glycerol + tween 80+ grape	Salmon Cold smoked Salmon Cold smoked wild pacific sockeye salmon	lipid oxidation Retarded lipid oxidation Microbial growth inhibited by olive leaf extracts Effective against L. monocytogenes Improved shelf life by Deferring	(Castro et al., 2019) (Albertos et al., 2017) (Benabbou et al., 2018)
starch coatings (C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate Gelatin Chitosan (94.7% DD) 2 % chitosan (75-85 % DD) + gelatin	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract Olive leaf extract+glycerol Bacteriocin (divergicin M35) Glycerol + tween 80+ grape seed extract + Ziziphora essential oil	Salmon Cold smoked Salmon Cold smoked wild pacific sockeye salmon Rainbow trout	lipid oxidation Retarded lipid oxidation Microbial growth inhibited by olive leaf extracts Effective against L. monocytogenes Improved shelf life by Deferring microbial growth and lipid oxidation	(Castro et al., 2019) (Albertos et al., 2017) (Benabbou et al., 2018) (Kakaei and Shahbazi, 2016)
starch coatings (C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate Gelatin Chitosan (94.7% DD) 2 % chitosan (75-85 % DD) + gelatin 1% agar + fish protein	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract Olive leaf extract+glycerol Bacteriocin (divergicin M35) Glycerol + tween 80+ grape seed extract + Ziziphora	Salmon Cold smoked Salmon Cold smoked wild pacific sockeye salmon Rainbow trout Flounder (Paralichthys	lipid oxidation Retarded lipid oxidation Microbial growth inhibited by olive leaf extracts Effective against L. monocytogenes Improved shelf life by Deferring microbial growth and lipid oxidation Improved shelf life by improving	(Castro et al., 2019) (Albertos et al., 2017) (Benabbou et al., 2018)
starch coatings (C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate Gelatin Chitosan (94.7% DD) 2 % chitosan (75-85 % DD) + gelatin 1% agar + fish protein	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract Olive leaf extract+glycerol Bacteriocin (divergicin M35) Glycerol + tween 80+ grape seed extract + Ziziphora essential oil	Salmon Cold smoked Salmon Cold smoked wild pacific sockeye salmon Rainbow trout	lipid oxidation Retarded lipid oxidation Microbial growth inhibited by olive leaf extracts Effective against L. monocytogenes Improved shelf life by Deferring microbial growth and lipid oxidation Improved shelf life by improving biochemical and microbiological	(Castro et al., 2019) (Albertos et al., 2017) (Benabbou et al., 2018) (Kakaei and Shahbazi, 2016)
starch coatings (C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate Gelatin Chitosan (94.7% DD) 2 % chitosan (75-85 %	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract Olive leaf extract+glycerol Bacteriocin (divergicin M35) Glycerol + tween 80+ grape seed extract + Ziziphora essential oil	Salmon Cold smoked Salmon Cold smoked wild pacific sockeye salmon Rainbow trout Flounder (Paralichthys	lipid oxidation Retarded lipid oxidation Microbial growth inhibited by olive leaf extracts Effective against L. monocytogenes Improved shelf life by Deferring microbial growth and lipid oxidation Improved shelf life by improving biochemical and microbiological parameter	(Castro et al., 2019) (Albertos et al., 2017) (Benabbou et al., 2018) (Kakaei and Shahbazi, 2016)
starch coatings (C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate Gelatin Chitosan (94.7% DD) 2 % chitosan (75-85 % DD) + gelatin 1% agar + fish protein hydrolysates	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract Olive leaf extract+glycerol Bacteriocin (divergicin M35) Glycerol + tween 80+ grape seed extract + Ziziphora essential oil	Salmon Cold smoked Salmon Cold smoked wild pacific sockeye salmon Rainbow trout Flounder (Paralichthys	lipid oxidation Retarded lipid oxidation Microbial growth inhibited by olive leaf extracts Effective against L. monocytogenes Improved shelf life by Deferring microbial growth and lipid oxidation Improved shelf life by improving biochemical and microbiological	(Castro et al., 2019) (Albertos et al., 2017) (Benabbou et al., 2018) (Kakaei and Shahbazi, 2016)
starch coatings (C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate Gelatin Chitosan (94.7% DD) 2 % chitosan (75-85 % DD) + gelatin 1% agar + fish protein hydrolysates 3% Chitosan + 3%	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract Olive leaf extract+glycerol Bacteriocin (divergicin M35) Glycerol + tween 80+ grape seed extract + Ziziphora essential oil Glycerol + Clove EOs Glycerol + Tween 80 with	Salmon Cold smoked Salmon Cold smoked wild pacific sockeye salmon Rainbow trout Flounder (Paralichthys orbignyanus)	lipid oxidation Retarded lipid oxidation Microbial growth inhibited by olive leaf extracts Effective against L. monocytogenes Improved shelf life by Deferring microbial growth and lipid oxidation Improved shelf life by improving biochemical and microbiological parameter	(Castro et al., 2019) (Albertos et al., 2017) (Benabbou et al., 2018) (Kakaei and Shahbazi, 2016) (da Rocha et al., 2018)
starch coatings (C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate Gelatin Chitosan (94.7% DD) 2 % chitosan (75-85 % DD) + gelatin 1% agar + fish protein hydrolysates 3% Chitosan + 3% Sarcoplasmic protein	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract Olive leaf extract+glycerol Bacteriocin (divergicin M35) Glycerol + tween 80+ grape seed extract + Ziziphora essential oil Glycerol + Clove EOs Glycerol + Tween 80 with ginger EOs	Salmon Cold smoked Salmon Cold smoked wild pacific sockeye salmon Rainbow trout Flounder (Paralichthys orbignyanus) Red sea bream	lipid oxidation Retarded lipid oxidation Microbial growth inhibited by olive leaf extracts Effective against L. monocytogenes Improved shelf life by Deferring microbial growth and lipid oxidation Improved shelf life by improving biochemical and microbiological parameter Extend the shelf life	(Castro et al., 2019) (Albertos et al., 2017) (Benabbou et al., 2018) (Kakaei and Shahbazi, 2016) (da Rocha et al., 2018) (Cai et al., 2020)
starch coatings (C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate Gelatin Chitosan (94.7% DD) 2 % chitosan (75-85 % DD) + gelatin 1% agar + fish protein hydrolysates 3% Chitosan + 3% Sarcoplasmic protein 1 % Chitosan + 0.5 %	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract Olive leaf extract+glycerol Bacteriocin (divergicin M35) Glycerol + tween 80+ grape seed extract + Ziziphora essential oil Glycerol + Clove EOs Glycerol + Tween 80 with ginger EOs Glycerol + pomegranate peel	Salmon Cold smoked Salmon Cold smoked wild pacific sockeye salmon Rainbow trout Flounder (Paralichthys orbignyanus)	lipid oxidation Retarded lipid oxidation Microbial growth inhibited by olive leaf extracts Effective against L. monocytogenes Improved shelf life by Deferring microbial growth and lipid oxidation Improved shelf life by improving biochemical and microbiological parameter Extend the shelf life Reduce microbial spoilage and	(Castro et al., 2019) (Albertos et al., 2017) (Benabbou et al., 2018) (Kakaei and Shahbazi, 2016) (da Rocha et al., 2018)
starch coatings (C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate Gelatin Chitosan (94.7% DD) 2 % chitosan (75-85 % DD) + gelatin 1% agar + fish protein hydrolysates 3% Chitosan + 3% Sarcoplasmic protein 1 % Chitosan + 0.5 % Locust bean gum	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract Olive leaf extract+glycerol Bacteriocin (divergicin M35) Glycerol + tween 80+ grape seed extract + Ziziphora essential oil Glycerol + Clove EOs Glycerol + Tween 80 with ginger EOs Glycerol + pomegranate peel extract	Salmon Cold smoked Salmon Cold smoked wild pacific sockeye salmon Rainbow trout Flounder (Paralichthys orbignyanus) Red sea bream White shrimps	lipid oxidation Retarded lipid oxidation Microbial growth inhibited by olive leaf extracts Effective against L. monocytogenes Improved shelf life by Deferring microbial growth and lipid oxidation Improved shelf life by improving biochemical and microbiological parameter Extend the shelf life Reduce microbial spoilage and volatile bases production	(Castro et al., 2019) (Albertos et al., 2017) (Benabbou et al., 2018) (Kakaei and Shahbazi, 2016) (da Rocha et al., 2018) (Cai et al., 2020) (Licciardello et al., 2018)
starch coatings (C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate Gelatin Chitosan (94.7% DD) 2 % chitosan (75-85 % DD) + gelatin 1% agar + fish protein hydrolysates 3% Chitosan + 3% Sarcoplasmic protein 1 % Chitosan + 0.5 % Locust bean gum	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract Olive leaf extract+glycerol Bacteriocin (divergicin M35) Glycerol + tween 80+ grape seed extract + Ziziphora essential oil Glycerol + Clove EOs Glycerol + Tween 80 with ginger EOs Glycerol + pomegranate peel	Salmon Cold smoked Salmon Cold smoked wild pacific sockeye salmon Rainbow trout Flounder (Paralichthys orbignyanus) Red sea bream	lipid oxidation Retarded lipid oxidation Microbial growth inhibited by olive leaf extracts Effective against L. monocytogenes Improved shelf life by Deferring microbial growth and lipid oxidation Improved shelf life by improving biochemical and microbiological parameter Extend the shelf life Reduce microbial spoilage and	(Castro et al., 2019) (Albertos et al., 2017) (Benabbou et al., 2018) (Kakaei and Shahbazi, 2016) (da Rocha et al., 2018) (Cai et al., 2020)
starch coatings (C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate Gelatin Chitosan (94.7% DD) 2 % chitosan (75-85 % DD) + gelatin 1% agar + fish protein	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract Olive leaf extract+glycerol Bacteriocin (divergicin M35) Glycerol + tween 80+ grape seed extract + Ziziphora essential oil Glycerol + Clove EOs Glycerol + Tween 80 with ginger EOs Glycerol + pomegranate peel extract	Salmon Cold smoked Salmon Cold smoked wild pacific sockeye salmon Rainbow trout Flounder (Paralichthys orbignyanus) Red sea bream White shrimps	lipid oxidation Retarded lipid oxidation Microbial growth inhibited by olive leaf extracts Effective against L. monocytogenes Improved shelf life by Deferring microbial growth and lipid oxidation Improved shelf life by improving biochemical and microbiological parameter Extend the shelf life Reduce microbial spoilage and volatile bases production Reduced oil uptake and moisture loss	(Castro et al., 2019) (Albertos et al., 2017) (Benabbou et al., 2018) (Kakaei and Shahbazi, 2016) (da Rocha et al., 2018) (Cai et al., 2020) (Licciardello et al., 2018)
starch coatings (C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate Gelatin Chitosan (94.7% DD) 2 % chitosan (75-85 % DD) + gelatin 1% agar + fish protein hydrolysates 3% Chitosan + 3% Sarcoplasmic protein 1 % Chitosan + 0.5 % Locust bean gum 1 % Basil seeds gum	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract Olive leaf extract+glycerol Bacteriocin (divergicin M35) Glycerol + tween 80+ grape seed extract + Ziziphora essential oil Glycerol + Clove EOs Glycerol + Tween 80 with ginger EOs Glycerol + pomegranate peel extract Glycerol + (99.5%) Thymol	Salmon Cold smoked Salmon Cold smoked wild pacific sockeye salmon Rainbow trout Flounder (Paralichthys orbignyanus) Red sea bream White shrimps Pacific white shrimp	lipid oxidation Retarded lipid oxidation Microbial growth inhibited by olive leaf extracts Effective against L. monocytogenes Improved shelf life by Deferring microbial growth and lipid oxidation Improved shelf life by improving biochemical and microbiological parameter Extend the shelf life Reduce microbial spoilage and volatile bases production Reduced oil uptake and moisture loss during fryig	<ul> <li>(Castro et al., 2019)</li> <li>(Albertos et al., 2017)</li> <li>(Benabbou et al., 2018)</li> <li>(Kakaei and Shahbazi, 2016)</li> <li>(da Rocha et al., 2018)</li> <li>(Cai et al., 2020)</li> <li>(Licciardello et al., 2018)</li> <li>(Khazaei et al., 2016)</li> </ul>
starch coatings (C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate Gelatin Chitosan (94.7% DD) 2 % chitosan (75-85 % DD) + gelatin 1% agar + fish protein nydrolysates 3% Chitosan + 3% Sarcoplasmic protein 1 % Gaircoplasmic protein 1 % Chitosan + 0.5 % Locust bean gum 1 % Basil seeds gum	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract Olive leaf extract+glycerol Bacteriocin (divergicin M35) Glycerol + tween 80+ grape seed extract + Ziziphora essential oil Glycerol + Clove EOs Glycerol + Tween 80 with ginger EOs Glycerol + pomegranate peel extract Glycerol + (99.5%) Thymol Glycerol + Tween 80 + EOs	Salmon Cold smoked Salmon Cold smoked wild pacific sockeye salmon Rainbow trout Flounder (Paralichthys orbignyanus) Red sea bream White shrimps	lipid oxidation Retarded lipid oxidation Microbial growth inhibited by olive leaf extracts Effective against L. monocytogenes Improved shelf life by Deferring microbial growth and lipid oxidation Improved shelf life by improving biochemical and microbiological parameter Extend the shelf life Reduce microbial spoilage and volatile bases production Reduced oil uptake and moisture loss during fryig reduce melanosis prevent microbial	(Castro et al., 2019) (Albertos et al., 2017) (Benabbou et al., 2018) (Kakaei and Shahbazi, 2016) (da Rocha et al., 2018) (Cai et al., 2020) (Licciardello et al., 2018)
starch coatings (C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate Gelatin Chitosan (94.7% DD) 2 % chitosan (75-85 % DD) + gelatin 1% agar + fish protein hydrolysates 3% Chitosan + 3% Sarcoplasmic protein 1 % Chitosan + 0.5 % Locust bean gum	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract Olive leaf extract+glycerol Bacteriocin (divergicin M35) Glycerol + tween 80+ grape seed extract + Ziziphora essential oil Glycerol + Clove EOs Glycerol + Tween 80 with ginger EOs Glycerol + pomegranate peel extract Glycerol + (99.5%) Thymol	Salmon Cold smoked Salmon Cold smoked wild pacific sockeye salmon Rainbow trout Flounder (Paralichthys orbignyanus) Red sea bream White shrimps Pacific white shrimp	lipid oxidation Retarded lipid oxidation Microbial growth inhibited by olive leaf extracts Effective against L. monocytogenes Improved shelf life by Deferring microbial growth and lipid oxidation Improved shelf life by improving biochemical and microbiological parameter Extend the shelf life Reduce microbial spoilage and volatile bases production Reduced oil uptake and moisture loss during fryig	<ul> <li>(Castro et al., 2019)</li> <li>(Albertos et al., 2017)</li> <li>(Benabbou et al., 2018)</li> <li>(Kakaei and Shahbazi, 2016)</li> <li>(da Rocha et al., 2018)</li> <li>(Cai et al., 2020)</li> <li>(Licciardello et al., 2018)</li> <li>(Khazaei et al., 2016)</li> </ul>
starch coatings (C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate Gelatin Chitosan (94.7% DD) 2 % chitosan (75-85 % DD) + gelatin 1% agar + fish protein hydrolysates 3% Chitosan + 3% Sarcoplasmic protein 1 % Chitosan + 0.5 % Locust bean gum 1 % Basil seeds gum Sweet potato starch	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract Olive leaf extract+glycerol Bacteriocin (divergicin M35) Glycerol + tween 80+ grape seed extract + Ziziphora essential oil Glycerol + Clove EOs Glycerol + Tween 80 with ginger EOs Glycerol + pomegranate peel extract Glycerol + (99.5%) Thymol Glycerol + Tween 80 + EOs (Thyme)	Salmon Cold smoked Salmon Cold smoked wild pacific sockeye salmon Rainbow trout Flounder (Paralichthys orbignyanus) Red sea bream White shrimps Pacific white shrimp Shrimp	lipid oxidation Retarded lipid oxidation Microbial growth inhibited by olive leaf extracts Effective against L. monocytogenes Improved shelf life by Deferring microbial growth and lipid oxidation Improved shelf life by improving biochemical and microbiological parameter Extend the shelf life Reduce microbial spoilage and volatile bases production Reduced oil uptake and moisture loss during fryig reduce melanosis prevent microbial growth and reduce lipid oxidation	<ul> <li>(Castro et al., 2019)</li> <li>(Albertos et al., 2017)</li> <li>(Benabbou et al., 2018)</li> <li>(Kakaei and Shahbazi, 2016)</li> <li>(da Rocha et al., 2018)</li> <li>(Cai et al., 2020)</li> <li>(Licciardello et al., 2018)</li> <li>(Khazaei et al., 2016)</li> <li>(Alotaibi and Tahergorabi, 2018)</li> </ul>
starch coatings (C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate Gelatin Chitosan (94.7% DD) 2 % chitosan (75-85 % DD) + gelatin 1% agar + fish protein hydrolysates 3% Chitosan + 3% Sarcoplasmic protein 1 % Chitosan + 0.5 % Locust bean gum 1 % Basil seeds gum Sweet potato starch	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract Olive leaf extract+glycerol Bacteriocin (divergicin M35) Glycerol + tween 80+ grape seed extract + Ziziphora essential oil Glycerol + Tween 80 with ginger EOs Glycerol + Tween 80 with ginger EOs Glycerol + pomegranate peel extract Glycerol + Tween 80 + EOs (Thyme) 1% Protein (42.7%)- lipid	Salmon Cold smoked Salmon Cold smoked wild pacific sockeye salmon Rainbow trout Flounder (Paralichthys orbignyanus) Red sea bream White shrimps Pacific white shrimp	lipid oxidation Retarded lipid oxidation Microbial growth inhibited by olive leaf extracts Effective against L. monocytogenes Improved shelf life by Deferring microbial growth and lipid oxidation Improved shelf life by improving biochemical and microbiological parameter Extend the shelf life Reduce microbial spoilage and volatile bases production Reduced oil uptake and moisture loss during fryig reduce melanosis prevent microbial growth and reduce lipid oxidation Delayed the onset of melanosis and	<ul> <li>(Castro et al., 2019)</li> <li>(Albertos et al., 2017)</li> <li>(Benabbou et al., 2018)</li> <li>(Kakaei and Shahbazi, 2016)</li> <li>(da Rocha et al., 2018)</li> <li>(Cai et al., 2020)</li> <li>(Licciardello et al., 2018)</li> <li>(Khazaei et al., 2016)</li> </ul>
starch coatings (C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate Gelatin Chitosan (94.7% DD) 2 % chitosan (75-85 % DD) + gelatin 1% agar + fish protein nydrolysates 3% Chitosan + 3% Sarcoplasmic protein 1 % Chitosan + 0.5 % Locust bean gum 1 % Basil seeds gum Sweet potato starch 7% Chitosan (75.5% DD)	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract Olive leaf extract+glycerol Bacteriocin (divergicin M35) Glycerol + tween 80+ grape seed extract + Ziziphora essential oil Glycerol + Clove EOs Glycerol + Tween 80 with ginger EOs Glycerol + pomegranate peel extract Glycerol + Tween 80 + EOs (Thyme) 1% Protein (42.7%)- lipid (11.48%) concentrate	Salmon Cold smoked Salmon Cold smoked wild pacific sockeye salmon Rainbow trout Flounder (Paralichthys orbignyanus) Red sea bream White shrimps Pacific white shrimp Shrimp	lipid oxidation Retarded lipid oxidation Microbial growth inhibited by olive leaf extracts Effective against L. monocytogenes Improved shelf life by Deferring microbial growth and lipid oxidation Improved shelf life by improving biochemical and microbiological parameter Extend the shelf life Reduce microbial spoilage and volatile bases production Reduced oil uptake and moisture loss during fryig reduce melanosis prevent microbial growth and reduce lipid oxidation Delayed the onset of melanosis and maintained sensory qualities	<ul> <li>(Castro et al., 2019)</li> <li>(Albertos et al., 2017)</li> <li>(Benabbou et al., 2018)</li> <li>(Kakaei and Shahbazi, 2016)</li> <li>(Kakaei and Shahbazi, 2016)</li> <li>(da Rocha et al., 2018)</li> <li>(Cai et al., 2020)</li> <li>(Licciardello et al., 2018)</li> <li>(Khazaei et al., 2016)</li> <li>(Alotaibi and Tahergorabi, 2018)</li> <li>(Arancibia et al., 2015)</li> </ul>
starch coatings (C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate Gelatin Chitosan (94.7% DD) 2 % chitosan (75-85 % DD) + gelatin 1% agar + fish protein nydrolysates 3% Chitosan + 3% Sarcoplasmic protein 1 % Chitosan + 0.5 % Locust bean gum 1 % Basil seeds gum Sweet potato starch 7% Chitosan (75.5% DD)	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract Olive leaf extract+glycerol Bacteriocin (divergicin M35) Glycerol + tween 80+ grape seed extract + Ziziphora essential oil Glycerol + Tween 80 with ginger EOs Glycerol + Tween 80 with ginger EOs Glycerol + pomegranate peel extract Glycerol + Tween 80 + EOs (Thyme) 1% Protein (42.7%)- lipid	Salmon Cold smoked Salmon Cold smoked wild pacific sockeye salmon Rainbow trout Flounder (Paralichthys orbignyanus) Red sea bream White shrimps Pacific white shrimp Shrimp	lipid oxidation Retarded lipid oxidation Microbial growth inhibited by olive leaf extracts Effective against L. monocytogenes Improved shelf life by Deferring microbial growth and lipid oxidation Improved shelf life by improving biochemical and microbiological parameter Extend the shelf life Reduce microbial spoilage and volatile bases production Reduced oil uptake and moisture loss during fryig reduce melanosis prevent microbial growth and reduce lipid oxidation Delayed the onset of melanosis and	<ul> <li>(Castro et al., 2019)</li> <li>(Albertos et al., 2017)</li> <li>(Benabbou et al., 2018)</li> <li>(Kakaei and Shahbazi, 2016)</li> <li>(da Rocha et al., 2018)</li> <li>(Cai et al., 2020)</li> <li>(Licciardello et al., 2018)</li> <li>(Khazaei et al., 2016)</li> <li>(Alotaibi and Tahergorabi, 2018)</li> </ul>
starch coatings (C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate Gelatin Chitosan (94.7% DD) 2 % chitosan (75-85 % DD) + gelatin 1% agar + fish protein nydrolysates 3% Chitosan + 3% Sarcoplasmic protein 1 % Chitosan + 0.5 % Locust bean gum 1 % Basil seeds gum Sweet potato starch 7% Chitosan (75.5% DD)	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract Olive leaf extract+glycerol Bacteriocin (divergicin M35) Glycerol + tween 80+ grape seed extract + Ziziphora essential oil Glycerol + Clove EOs Glycerol + Tween 80 with ginger EOs Glycerol + pomegranate peel extract Glycerol + Tween 80 + EOs (Thyme) 1% Protein (42.7%)- lipid (11.48%) concentrate	Salmon Cold smoked Salmon Cold smoked wild pacific sockeye salmon Rainbow trout Flounder (Paralichthys orbignyanus) Red sea bream White shrimps Pacific white shrimp Shrimp	lipid oxidation Retarded lipid oxidation Microbial growth inhibited by olive leaf extracts Effective against L. monocytogenes Improved shelf life by Deferring microbial growth and lipid oxidation Improved shelf life by improving biochemical and microbiological parameter Extend the shelf life Reduce microbial spoilage and volatile bases production Reduced oil uptake and moisture loss during fryig reduce melanosis prevent microbial growth and reduce lipid oxidation Delayed the onset of melanosis and maintained sensory qualities Reduced weight loss and improved	<ul> <li>(Castro et al., 2019)</li> <li>(Albertos et al., 2017)</li> <li>(Benabbou et al., 2018)</li> <li>(Kakaei and Shahbazi, 2016)</li> <li>(Kakaei and Shahbazi, 2016)</li> <li>(da Rocha et al., 2018)</li> <li>(Cai et al., 2020)</li> <li>(Licciardello et al., 2018)</li> <li>(Khazaei et al., 2016)</li> <li>(Alotaibi and Tahergorabi, 2018)</li> <li>(Arancibia et al., 2015)</li> </ul>
starch coatings (C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate Gelatin Chitosan (94.7% DD) 2 % chitosan (75-85 % DD) + gelatin 1% agar + fish protein 1% agar + fish protein 1% dgar + fish protein 1% Garcoplasmic protein 1 % Chitosan + 3% Sarcoplasmic protein 1 % Basil seeds gum Sweet potato starch 7% Chitosan (75.5% DD) 1% Chitosan (85% DD)	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract Olive leaf extract+glycerol Bacteriocin (divergicin M35) Glycerol + tween 80+ grape seed extract + Ziziphora essential oil Glycerol + Clove EOs Glycerol + Tween 80 with ginger EOs Glycerol + pomegranate peel extract Glycerol + Tween 80 + EOs (Thyme) 1% Protein (42.7%)- lipid (11.48%) concentrate 3% gelatin	Salmon Cold smoked Salmon Cold smoked wild pacific sockeye salmon Rainbow trout Flounder (Paralichthys orbignyanus) Red sea bream White shrimps Pacific white shrimp Shrimp Shrimp	lipid oxidation Retarded lipid oxidation Microbial growth inhibited by olive leaf extracts Effective against L. monocytogenes Improved shelf life by Deferring microbial growth and lipid oxidation Improved shelf life by improving biochemical and microbiological parameter Extend the shelf life Reduce microbial spoilage and volatile bases production Reduced oil uptake and moisture loss during fryig reduce melanosis prevent microbial growth and reduce lipid oxidation Delayed the onset of melanosis and maintained sensory qualities Reduced weight loss and improved texture and color changes	<ul> <li>(Castro et al., 2019)</li> <li>(Albertos et al., 2017)</li> <li>(Benabbou et al., 2018)</li> <li>(Kakaei and Shahbazi, 2016)</li> <li>(da Rocha et al., 2018)</li> <li>(Cai et al., 2020)</li> <li>(Licciardello et al., 2018)</li> <li>(Khazaei et al., 2016)</li> <li>(Alotaibi and Tahergorabi, 2018)</li> <li>(Arancibia et al., 2015)</li> <li>(Farajzadeh et al., 2016)</li> </ul>
starch coatings (C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate Gelatin Chitosan (94.7% DD) 2 % chitosan (75-85 % DD) + gelatin 1% agar + fish protein 1% agar + fish protein 1% dgar + fish protein 1% Garcoplasmic protein 1 % Chitosan + 3% Sarcoplasmic protein 1 % Basil seeds gum Sweet potato starch 7% Chitosan (75.5% DD) 1% Chitosan (85% DD)	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract Olive leaf extract+glycerol Bacteriocin (divergicin M35) Glycerol + tween 80+ grape seed extract + Ziziphora essential oil Glycerol + Clove EOs Glycerol + Tween 80 with ginger EOs Glycerol + pomegranate peel extract Glycerol + Tween 80 + EOs (Thyme) 1% Protein (42.7%)- lipid (11.48%) concentrate	Salmon Cold smoked Salmon Cold smoked wild pacific sockeye salmon Rainbow trout Flounder (Paralichthys orbignyanus) Red sea bream White shrimps Pacific white shrimp Shrimp	lipid oxidation Retarded lipid oxidation Microbial growth inhibited by olive leaf extracts Effective against L. monocytogenes Improved shelf life by Deferring microbial growth and lipid oxidation Improved shelf life by improving biochemical and microbiological parameter Extend the shelf life Reduce microbial spoilage and volatile bases production Reduced oil uptake and moisture loss during fryig reduce melanosis prevent microbial growth and reduce lipid oxidation Delayed the onset of melanosis and maintained sensory qualities Reduced weight loss and improved texture and color changes Inhibited melanosis and improved the	<ul> <li>(Castro et al., 2019)</li> <li>(Albertos et al., 2017)</li> <li>(Benabbou et al., 2018)</li> <li>(Kakaei and Shahbazi, 2016)</li> <li>(Kakaei and Shahbazi, 2016)</li> <li>(da Rocha et al., 2018)</li> <li>(Cai et al., 2020)</li> <li>(Licciardello et al., 2018)</li> <li>(Khazaei et al., 2016)</li> <li>(Alotaibi and Tahergorabi, 2018)</li> <li>(Arancibia et al., 2015)</li> </ul>
starch coatings (C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate Gelatin Chitosan (94.7% DD) 2 % chitosan (75-85 % DD) + gelatin 1% agar + fish protein 1% dagar + fish protein 1% Chitosan + 3% Sarcoplasmic protein 1 % Chitosan + 0.5 % Locust bean gum 1 % Basil seeds gum Sweet potato starch 7% Chitosan (75.5% DD) 1% Chitosan (85% DD) Chitosan (90% DD)	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract Olive leaf extract+glycerol Bacteriocin (divergicin M35) Glycerol + tween 80+ grape seed extract + Ziziphora essential oil Glycerol + Clove EOs Glycerol + Tween 80 with ginger EOs Glycerol + pomegranate peel extract Glycerol + (99.5%) Thymol Glycerol + Tween 80 + EOs (Thyme) 1% Protein (42.7%)- lipid (11.48%) concentrate 3% gelatin Pomegranate peel extract	Salmon Cold smoked Salmon Cold smoked wild pacific sockeye salmon Rainbow trout Flounder (Paralichthys orbignyanus) Red sea bream White shrimps Pacific white shrimp Shrimp Shrimp Pacific white shrimp	lipid oxidation Retarded lipid oxidation Microbial growth inhibited by olive leaf extracts Effective against L. monocytogenes Improved shelf life by Deferring microbial growth and lipid oxidation Improved shelf life by improving biochemical and microbiological parameter Extend the shelf life Reduce microbial spoilage and volatile bases production Reduced oil uptake and moisture loss during fryig reduce melanosis prevent microbial growth and reduce lipid oxidation Delayed the onset of melanosis and maintained sensory qualities Reduced weight loss and improved texture and color changes Inhibited melanosis and improved the sensory qualities	<ul> <li>(Castro et al., 2019)</li> <li>(Albertos et al., 2017)</li> <li>(Benabbou et al., 2018)</li> <li>(Kakaei and Shahbazi, 2016)</li> <li>(da Rocha et al., 2018)</li> <li>(Cai et al., 2020)</li> <li>(Licciardello et al., 2018)</li> <li>(Khazaei et al., 2016)</li> <li>(Alotaibi and Tahergorabi, 2018)</li> <li>(Arancibia et al., 2015)</li> <li>(Farajzadeh et al., 2015)</li> </ul>
starch coatings (C) Edible Packaging for Sea 1.25 % chitosan Whey protein concentrate Gelatin Chitosan (94.7% DD) 2 % chitosan (75-85 % DD) + gelatin 1% agar + fish protein 1% agar + fish protein 1% dgar + fish protein 1% Garcoplasmic protein 1 % Chitosan + 3% Sarcoplasmic protein 1 % Basil seeds gum Sweet potato starch 7% Chitosan (75.5% DD) 1% Chitosan (85% DD)	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract Olive leaf extract+glycerol Bacteriocin (divergicin M35) Glycerol + tween 80+ grape seed extract + Ziziphora essential oil Glycerol + Clove EOs Glycerol + Tween 80 with ginger EOs Glycerol + pomegranate peel extract Glycerol + Tween 80 + EOs (Thyme) 1% Protein (42.7%)- lipid (11.48%) concentrate 3% gelatin	Salmon Cold smoked Salmon Cold smoked wild pacific sockeye salmon Rainbow trout Flounder (Paralichthys orbignyanus) Red sea bream White shrimps Pacific white shrimp Shrimp Shrimp	lipid oxidation Retarded lipid oxidation Microbial growth inhibited by olive leaf extracts Effective against L. monocytogenes Improved shelf life by Deferring microbial growth and lipid oxidation Improved shelf life by improving biochemical and microbiological parameter Extend the shelf life Reduce microbial spoilage and volatile bases production Reduced oil uptake and moisture loss during fryig reduce melanosis prevent microbial growth and reduce lipid oxidation Delayed the onset of melanosis and maintained sensory qualities Reduced weight loss and improved texture and color changes Inhibited melanosis and improved the	<ul> <li>(Castro et al., 2019)</li> <li>(Albertos et al., 2017)</li> <li>(Benabbou et al., 2018)</li> <li>(Kakaei and Shahbazi, 2016)</li> <li>(da Rocha et al., 2018)</li> <li>(Cai et al., 2020)</li> <li>(Licciardello et al., 2018)</li> <li>(Khazaei et al., 2016)</li> <li>(Alotaibi and Tahergorabi, 2018)</li> <li>(Arancibia et al., 2015)</li> <li>(Farajzadeh et al., 2016)</li> </ul>
<ul> <li>starch coatings</li> <li>(C) Edible Packaging for Sea</li> <li>(C) Editosan (75-85 %</li> <li>(C) Editosan (75-5% DD)</li> <li>(C) Editosan (90% DD)</li> <li>(C) Editosan (90% DD)</li> </ul>	Foods and Meat Products 0.8% glycerol+ Tween 80+grape seed extract+carvacrol Glycerol+green tea extract Olive leaf extract+glycerol Bacteriocin (divergicin M35) Glycerol + tween 80+ grape seed extract + Ziziphora essential oil Glycerol + Clove EOs Glycerol + Tween 80 with ginger EOs Glycerol + pomegranate peel extract Glycerol + (99.5%) Thymol Glycerol + Tween 80 + EOs (Thyme) 1% Protein (42.7%)- lipid (11.48%) concentrate 3% gelatin Pomegranate peel extract	Salmon Cold smoked Salmon Cold smoked wild pacific sockeye salmon Rainbow trout Flounder (Paralichthys orbignyanus) Red sea bream White shrimps Pacific white shrimp Shrimp Shrimp Pacific white shrimp	lipid oxidation Retarded lipid oxidation Microbial growth inhibited by olive leaf extracts Effective against L. monocytogenes Improved shelf life by Deferring microbial growth and lipid oxidation Improved shelf life by improving biochemical and microbiological parameter Extend the shelf life Reduce microbial spoilage and volatile bases production Reduced oil uptake and moisture loss during fryig reduce melanosis prevent microbial growth and reduce lipid oxidation Delayed the onset of melanosis and maintained sensory qualities Reduced weight loss and improved texture and color changes Inhibited melanosis and improved the sensory qualities	<ul> <li>(Castro et al., 2019)</li> <li>(Albertos et al., 2017)</li> <li>(Benabbou et al., 2018)</li> <li>(Kakaei and Shahbazi, 2016)</li> <li>(da Rocha et al., 2018)</li> <li>(Cai et al., 2020)</li> <li>(Licciardello et al., 2018)</li> <li>(Khazaei et al., 2016)</li> <li>(Alotaibi and Tahergorabi, 2018)</li> <li>(Arancibia et al., 2015)</li> <li>(Farajzadeh et al., 2015)</li> </ul>

Table 3 (continued)

Edible Film/Coating	Additives	Food Commodities	Significant Function	References
0.5 and 1.5% Chitosan (91% DD)		Atlantic salmon	1.5% Chitosan maintained better quality and controlled microbial growth	(Soares et al., 2015)
4% Collagen	0.1, 0.3, 0.5, and 0.7% Lysozyme + 1% glycerol	Salmon	Reduced weight loss and improved texture and sensory qualities	(Wang et al., 2017)
1% Carrageenan	1% EOs (lemon)	Rainbow trout	Preserved physical–chemical, morphological, and olfactory characteristics	(Volpe et al., 2019)
1% Alginate	0.3% Tannic acid, 0.3% quebracho tannin, and 1% ascorbic acid	Rainbow trout	Reduced microbial counts and lipid oxidation	(Sáez et al., 2020)
Whey protein concentrate	Glycerol	Rainbow trout	Suppressed microbial growth and enhanced sensory attributes	(Oğuzhan Yıldız & Yangılar, 2016)
Whey protein	Glycerol+NaOH	Atlantic salmon	Improved the overall quality of salmon	(Rodriguez-Turienzo, Cobos and Diaz, 2012)
Caseiante	Ascorbic acid	Beef	Effect of gamma irradiation on microbiological characteristic of ground beef	(Hassan et al., 2018)

Abbreviations: DD: de-acetylated, HPMC: Hydroxypropyl methylcellulose, EO: Essential oil.

#### 4.1. Fruits and vegetables

Consumers always prefer to choose fruits and vegetables in fresh form, which has led to the continuous development of advanced methods that helps in maintaining quality and shelf life of the produce (Flores-López et al., 2016). Nevertheless, high moisture and microbial deterioration make these products highly perishable and limit their storability. Applying adequate packaging such as edible packaging can enhance its shelf life by creating a barrier against microbes, moisture, and gases (Pizato et al., 2019). A number of works have been reported based on bio-polymer application in fruits and vegetables (Table 3 (A&B)). For example, a chitosan-based film in combination with  $TiO_2$ has experimented for the storage of grapes (Zhang et al., 2017), application of alginate-based composite film on shelf life of fresh fig was studied by Reyes-Avalos et al., (2016), the effect of beeswax content on hydroxypropyl methylcellulose-based edible film on postharvest quality of coated plums was evaluated by Navarro-Tarazaga et al., (2011).

Edible film/coatings can improve the quality and shelf life of various fruits by inhibiting lipid oxidation, delaying moisture loss, preventing discoloration, and maintaining the fruits' appearance during marketing by minimizing dirt and dust contact, entrapping volatile flavor compounds, and acting as carriers of food additives such as antimicrobial and antioxidative agents. Ochoa et al. (2011) successfully improved the quality and shelf life of delicious golden apples by applying edible layers comprised of natural wax extracted from Euphorbia antisyphilitica and potent antioxidant 0.01% ellagic acid (EA). Walnuts and pine nuts coated with a homogenized coating solution of whey protein isolate with carnauba wax exhibited lower oxidative and hydrolytic rancidity and improved sensory quality (Mehyar et al., 2012). Fagundes et al., (2015), evaluated the beneficial properties of composite edible films formulated with HPMC, beeswax, and different food preservatives having an antifungal property like sodium benzoate, sodium ethyl paraben, and sodium methyl paraben applied to cherry tomato with artificially inoculated black spot fungi Alternaria alternata during cold storage. The authors observed that sodium benzoate (2%) based edible packaging significantly reduced weight loss, respiration rate, and maintained the firmness of the cherry tomatoes.

#### 4.2. Dairy products

Milk and milk products are considered a good source of dietary supplements for better health of both children and adults (Cardador & Gallego, 2016). The literature survey shows that cheese packaging is considered one of the potential application areas of the edible film (V et al, 2021). Packaging, especially with antimicrobial film in cheese observed to have considerable influences on its shelf life. Fajardo et al. (2010) evaluated the efficacy of chitosan-based film as a carrier of natamycin to improve storage stability of Saloio cheese, and observed that the product was stable until seven days at ambient storage conditions. Mahcene et al. (2020) assessed the preservative effect of sodium alginate-based edible film incorporated with essential oil on homemade cheese, and they concluded that edible packaging is an effective preservation method of cheese.

Martins et al. (2010) evaluated the shelf-life extension of Ricotta cheese, a soft Italian cheese, during cold storage (4°C) upon using galactomannan-based edible coatings in combination with nisin (50 IU g–1) against Listeria monocytogenes. Authors found that edible coatings with nisin delay the microbial growth significantly (P<0.05), increased the tensile strength (0.84 to 1.46 MPa), increased the opacity (3.68 to 4.59%), improved the elongation breakpoint (50.93 to 68.16%), increased the CO<sub>2</sub> permeability (1.96 to  $6.31 \times 10^{12}$ ) cm<sup>3</sup>. (Pa.s.m)<sup>-1</sup>), and decreased the O<sub>2</sub> permeability (1.84 to  $1.35 \times 10^{-12}$ ) cm<sup>3</sup> x (Pa.s.m)<sup>-1</sup>). Similarly, whey protein (10% w/w) based coating in combination with chitooligosaccharide (20g L<sup>-1</sup>) and lactic acid (6g L<sup>-1</sup>) potentially inhibited the microbial growth (<2.0 cfu/g) on laboratory manufactured cheese after 60 days long storage at 10°C (Ramos et al., 2012).

#### 4.3. Meat and meat products

Meat and meat products are considered favorable food among consumers due to their unique taste and nutritional benefits. Nevertheless, meat is highly susceptible to spoilage due to microbial and chemical changes; therefore, its preservation requires special attention. Advanced packaging techniques such as intelligent and antimicrobial packaging have emerged as food safety hurdles technology primarily for the products like meat (Soni et al., 2018). The edible packaging concept is gaining much popularity among meat and meat products due to its significant role in improving physicochemical and sensory properties (Galus & Kadzińska, 2015). Research progress in applying edible film in meat and meat products was briefly summarized in Table 3 (C).

Essential oils have become more widely used in edible films due to their antioxidant and antimicrobial properties in recent years. Moradi et al. (2016) assessed the antimicrobial effects of zein-based films in combination with *Zataria multiflora* Boiss. essential oil (ZEO) (3%) and monulaurin (1%) against *E. coli* O157: H7 and *Listeria monocytogenes in vitro* and minced beef. The authors found that ZEO significantly enhanced the antimicrobial activity of the film. Similarly, Ferulago Angulate Essential Oil (FAEO) (0.05%) incorporated with gelatin-chitosan-based film inhibited the microbial growth and improved the shelf life of turkey meat (Naseri et al., 2020).

### 4.4. Seafoods

Seafood, which includes fish and fish products, generally has a limited shelf life due to the rapid growth of microbes, which can pose a threat to consumers' health as well as result in economic loss. (Gómez-Estaca et al., 2010; López de Lacey et al., 2014). In recent years, the edible packaging concept is evolved as an efficient strategy to enhance the storage stability of fish (Günlü & Koyun, 2013). Successful application of several polysaccharides such as chitosan (Remya et al., 2016), sodium alginate-carboxy methylcellulose (Rezaei & Shahbazi, 2018), whey protein (Seyfzadeh et al 2013), etc., which various researchers have already explored. Some of the recent progress and their major findings are given in Table 3 (C).

The edible film/coatings, which are primarily chitosan-gelatin based, effectively preserve the desired quality, extend the shelf life, and improve the texture and color of sea and meat products by reducing spoilage, reducing the accumulation of volatile compounds and oxidation substances, and minimizing weight loss. Castro et al., (2019), verified the potential of whey protein concentrate film in combination with green tea extract applied on fresh salmon and found that their combination effectively delayed the lipid oxidation of fresh salmon until the fourteen days of storage. The incorporation of essential oils into edible films due to their antioxidant and antimicrobial activity has been more documented in the past few years. Ginger essential oil incorporated with fish sarcoplasmic protein and chitosan applied to red sea bream significantly reduced the oxidation, protected from microbial degradation, and extended shelf life of the red sea bream (Cai et al., 2020).

# 5. Safety and regulation for edible films

Food safety and regulation vary from country to country. According to EU and US regulations, edible film and coating can be considered food ingredients, additives, contact materials, or packaging materials. As a result, the constituents used for its formulation should have Generally Recommended As Safe (GRAS) status as per the regulations of the Food and Drug Administration (FDA) regulation (Dhall, 2013). Nevertheless, there are chances for the transformation of FFS into toxic substances due to changes that occur during the film development process (Giteru et al., 2020). Different cross-linking agents used for enhancing the film properties and interaction with gastrointestinal substances can also trigger the formation of toxic materials (Chiralt et al., 2018). In one study (Roşu et al., 2017), the effect of graphene oxide and its derivatives on the cytotoxicity of the methylcellulose-based film on human lungs was investigated, and lower toxicity was reported for reduced graphene oxide compared to graphene oxide. The effect of modification techniques and ingredient selection plays a critical role in edible film safety but is rarely reported in edible film studies. Moreover, the addition of nanomaterials in the film may cause several toxicological effects on human bodies, as explained in section 2.5. At present, there is no specific legislation for nanomaterial use in food packaging, and the recommendations differ by nation. As per the guidelines of the Institute of Food Science and Technology (IFST) in the United Kingdom, nanomaterials should be considered hazardous unless clear safety proof is available (Jeevahan et al., 2020). According to EU and Switzerland legislation, information on nanomaterial risk and/or legally binding definitions of nanomaterial need to be produced. Labeling the presence of nanomaterials in a specific film is critical for communicating risk elements to consumers. In EU definitions, size is used as an identifier for nanomaterials in regulatory purposes (Bizymis & Tzia, 2021). The USFDA publishes a list of food ingredients and contact substances and advises manufacturers to research and develop a toxicological profile for each container containing nanomaterial (USFDA 2014). Most countries, presently, do not have any regulations for the use of nanotechnology. More research on the nanomaterial's toxicological effects can help formulate new safety and regulations for its application.

# 6. Recent trends, challenges, and future perspectives in edible film

The edible film research area is constantly evolving, with new raw materials for film formation, active packaging development, nanotechnology applications, etc. All these efforts were targeted to develop biopolymers having properties par with conventional synthetic polymers and their economic production by utilizing agricultural byproducts as raw material. The development of active films has been identified as one of the primary focus areas in packaging research in the last decades, which enable the shelf-life extension of perishable fruits and vegetables by the addition of various antimicrobial and antioxidant components into their base polymer matrix (Deng et al., 2020; Nair et al., 2020; Orozco-Parra et al., 2020). Active edible films have active interaction with contained food and contribute health benefits to consumers (Moradi et al., 2021). For example, Orozco-Parra et al. (2020) developed a synbiotic film from cassava starch with the incorporation of insulin as a prebiotic molecule and L.casei as the probiotic bacteria. The developed film has shown decreased viability loss of probiotic bacteria during simulated gastric condition study.

Similarly, the successful use of edible film for the transport of probiotic bacteria was also reported by other authors (Ebrahimi et al., 2018; Soukoulis et al., 2017). In recent years food-processing byproducts-based edible films are gaining popularity. It allows the valorization of industrial by-products, and their low-cost helps develop edible film economically. By-products of fruits and vegetable processing, marine food processing, and edible oil processing industries proved the promising potential for the film preparations (Aloui et al., 2019; Benbettaïeb et al., 2019; Hromiš et al., 2022; Moghadam et al., 2020; Shroti and Saini, 2022 Valdés et al., 2020). As explained in the section 2.4 & 2.5, the edible films with tailored properties have been experimented with composite film formulation and nanotechnology application. Moreover, several attempts were also made to achieve this goal by cross-linking various biopolymers (Peng et al., 2021; Yerramathi et al., 2021; Zhang et al., 2022).

Despite all advantages, like all other newly developed technologies, edible films also face great challenges that need to be overcome to make them a commercial success. Even though a number of ways are experimented to improve the properties of the film to make it on par with petroleum-based polymers, poor mechanical properties, weak resistance against water and gases, and insufficient physical properties are the hurdle for its use in various food applications. Investigations were carried out to address these challenges by developing composite films and nanocomposite films. However, composite film production by multilayer approach tends to fail by delamination of layers, and time, energy, and cost requirement for the formation of the multilayer film are high. Another crucial property of films is their heat sealability. The range of optimum sealing temperature is narrow for biobased films. So, chances of undersealing and charring due to overheating are higher for these films. This negatively affects its applications in the formation of pouches and covers. The main reason restricting the commercial application of edible film technology is the inability to make large-sized films (>25cm), difficulties in maintaining the thickness, and a long-time requirement for drying (2-3 days). Application of nanotechnology is considered a trending area for edible films. Still, practical difficulties in isolation and homogeneous dispersion of nanomaterials in matrix polymers and economic aspects always create a challenge. Moreover, a lack of information on the toxicological effect of nanomaterial and other filmforming components generates fear and reluctance in consumers. Only limited studies were reported on the evaluation of how the aging of the film affects its properties. Since the primary intended use of packaging material is to contain the food and extend its shelf life during storage, thus this factor needs to be considered (Jeevahan et al., 2020). Among the reported research on edible film, emphasis on sensory analysis is relatively low. The sensory attributes of the film are the first and foremost important factor that decides the acceptance of the edible

packaging film. Ultimately consumer acceptance is the only deciding factor in changing our packaging concept. For achieving its commercial success, complete study and documentation are required to prove its biodegradability, organoleptic aspects, safety and security, and legal confirmation. In order to overcome these challenges, considerable research efforts need to concentrate on the following essential aspects. A detailed toxicological study of the film-forming components by emphasizing nanoparticles is required to be carried out. Any technology on a laboratory scale cannot benefit the consumers. So, the method for the production of the continuous edible film with consistent properties needs to be developed. Change in properties of edible film during aging and under different temperature and relative humidity conditions is also essential to confirm its utility in food storage purposes. Consumer awareness programs and advertisements of edible films can also increase their acceptability.

#### 7. Conclusion

Edible films have been identified as a healthy source of food protection from various elements, as they are naturally occurring, inexpensive, and renewable. The possibility of incorporating functional ingredients and excellent biodegradability further glorifies its attraction. Extensive research has been conducted to determine the best outcome and minimize drawbacks with new concepts such as composite film approach and nanotechnology application. More research on important aspects like property improvement, implementation of safety and regulation, exploration of new and economic sources, and commercial scale-up by continuous production is essential to its successful adoption.

# **Conflict of Interest Form**

The following authors have declared that there is no conflict of interest in publishing manuscript entitled "*Trends in Edible Packaging Films and its Prospective Future in Food: A Review*"

#### References

- Abral, H., Basri, A., Muhammad, F., Fernando, Y., Hafizulhaq, F., Mahardika, M., Sugiarti, E., Sapuan, S. M., Ilyas, R. A., & Stephane, I. (2019). A simple method for improving the properties of the sago starch films prepared by using ultrasonication treatment. *Food Hydrocolloids*, 93, 276–283. 10.1016/j.foodhyd.2019.02.012.
- Al-Hashimi, A. G., Ammar, A. B., Lakshmanan, G., Cacciola, F., & Lakhssassi, N. (2020). Development of a millet starch edible film containing clove essential oil. *Foods*, 9(2), 1–14. 10.3390/foods9020184.
- Albertos, I., Avena-Bustillos, R. J., Martín-Diana, A. B., Du, W. X., Rico, D., & McHugh, T. H. (2017). Antimicrobial Olive Leaf Gelatin films for enhancing the quality of cold-smoked Salmon. *Food Packaging and Shelf Life*, 13, 49–55. 10.1016/j.fpsl.2017.07.004.
- Alexandre, E. M. C., Lourenço, R. V., Bittante, A. M. Q. B., Moraes, I. C. F., & Sobral, P. J. do A. (2016). Gelatin-based films reinforced with montmorillonite and activated with nanoemulsion of ginger essential oil for food packaging applications. *Food Packaging* and Shelf Life, 10, 87–96. 10.1016/j.fpsl.2016.10.004.
- Alizadeh-Sani, M., Rhim, J.-W., Azizi-Lalabadi, M., Hemmati-Dinarvand, M., & Ehsani, A. (2020). Preparation and characterization of functional sodium caseinate/guar gum/TiO2/cumin essential oil composite film. *International Journal of Biological Macromolecules*, 145, 835–844. 10.1016/j.ijbiomac.2019.11.004.
- Alotaibi, S., & Tahergorabi, R. (2018). Development of a sweet potato starch-based coating and its effect on quality attributes of shrimp during refrigerated storage. *LWT*, 88, 203–209. 10.1016/j.lwt.2017.10.022.
- Aloui, H., Baraket, K., Sendon, R., Silva, A. S., & Khwaldia, K. (2019). Development and characterization of novel composite glycerol-plasticized films based on sodium caseinate and lipid fraction of tomato pomace by-product. *International Journal of Biological Macromolecules*, 139, 128–138. 10.1016/j.ijbiomac.2019.07.156.
- Alves, V. L. C. D., Rico, B. P. M., Cruz, M. S., Vicente, A. A., Khmelinskii, I., & Vieira, M. C. (2017). Preparation and characterization of a chitosan film with grape seed extract-carvacrol microcapsules and its effect on the shelf-life of refrigerated Salmon (Salmo salar). *LWT - Food Science and Technology*, 89, 525–534. 10.1016/j.lwt.2017.11.013.
- Ansorena, M. R., Zubeldía, F., & Marcovich, N. E. (2016). Active wheat gluten films obtained by thermoplastic processing. LWT - Food Science and Technology, 69, 47–54. 10.1016/j.lwt.2016.01.020.
- Arancibia, M. Y., López-Caballero, M. E., Gómez-Guillén, M. C., & Montero, P. (2015). Chitosan coatings enriched with active shrimp waste for shrimp preservation. *Food Control*, 54, 259–266. 10.1016/j.foodcont.2015.02.004.

- Askari, F., Sadeghi, E., Mohammadi, R., Rouhi, M., Taghizadeh, M., Hosein Shirgardoun, M., & Kariminejad, M. (2018). The physicochemical and structural properties of psyllium gum/modified starch composite edible film. *Journal of Food Processing and Preservation*, 42(10), 1–9. 10.1111/jfpp.13715.
- Bagheri, F., Radi, M., & Amiri, S. (2019). Drying conditions highly influence the characteristics of glycerol-plasticized alginate films. *Food Hydrocolloids*, 90, 162–171. 10.1016/j.foodhyd.2018.12.001.
- Baldwin, E. A., Burns, J. K., Kazokas, W., Brecht, J. K., Hagenmaier, R. D., Bender, R. J., & Pesis, E. (1999). Effect of two edible coatings with different permeability characteristics on mango (Mangifera indica L.) ripening during storage. *Postharvest Biology* and Technology, 17(3), 215–226. 10.1016/S0925-5214(99)00053-8.
- Bamdad, F., Goli, A. H., & Kadivar, M. (2006). Preparation and characterization of proteinous film from lentil (*Lens culinaris*): Edible film from lentil (*Lens culinaris*). Food Research International, 39(1), 106–111. 10.1016/j.foodres.2005.06.006.
- Belyamani, I., Prochazka, F., & Assezat, G. (2014). Production and characterization of sodium caseinate edible films made by blown-film extrusion. *Journal of Food Engineering*, 121(1), 39–47. 10.1016/j.jfoodeng.2013.08.019.
- Benabbou, R., Subirade, M., Desbiens, M., & Fliss, I. (2018). The impact of chitosandivergicin film on growth of listeria monocytogenes in Cold-smoked salmon. *Frontiers* in Microbiology, 9 NOV. 10.3389/fmicb.2018.02824.
- Benbettaïeb, N., O'Connell, C., Viaux, A., Bou-Maroun, E., Seuvre, A.-M., Brachais, C.-H., & Debeaufort, F. (2019). Sorption kinetic of aroma compounds by edible bio-based films from marine-by product macromolecules: Effect of relative humidity conditions. *Food Chemistry*, 298, Article 125064. 10.1016/j.foodchem.2019.125064.
- Bharti, S. K., Pathak, V., Arya, A., Alam, T., Rajkumar, V., & Verma, A. K. (2020). Packaging potential of Ipomoea batatas and κ-carrageenan biobased composite edible film: Its rheological, physicomechanical, barrier and optical characterization. *Journal of Food Processing and Preservation*, 1–11 *June*. 10.1111/jfpp.15153.
- Bhartiya, A., Aditya, J. P., Pal, R. S., Chandra, N., Kant, L., & Pattanayak, A. (2020). Bhat (Black soybean): A traditional legume with high nutritional and nutraceutical properties from nw himalayan region of india. *Indian Journal of Traditional Knowledge*, 19(2), 307–319.
- Bizymis, A.-P., & Tzia, C. (2021). Edible films and coatings: Properties for the selection of the components, evolution through composites and nanomaterials, and safety issues. *Critical Reviews in Food Science and Nutrition*, 1–16. 10.1080/10408398.2021.1934652.
- Bonilla, J., & Sobral, P. J. A. (2016). Investigation of the physicochemical, antimicrobial and antioxidant properties of gelatin-chitosan edible film mixed with plant ethanolic extracts. *Food Bioscience*, 16, 17–25. 10.1016/j.fbio.2016.07.003.
- Bourtoom, T. (2008). Edible films and coatings: Characteristics and properties. International Food Research Journal, 15(3), 237–248.
- Bourtoom, T. (2009). Edible protein films: Properties enhancement. International Food Research Journal, 16(1), 1–9.
- Bumbudsanpharoke, N., Choi, J., & Ko, S. (2015). Applications of nanomaterials in food packaging. Journal of Nanoscience and Nanotechnology, 15(9), 6357–6372. 10.1166/jnn.2015.10847.
- Cai, L., Wang, Y., & Cao, A. (2020). The physiochemical and preservation properties of fish sarcoplasmic protein/chitosan composite films containing ginger essential oil emulsions. *Journal of Food Process Engineering*, (10), 43. 10.1111/jfpe.13495.
- Çağrı Mehmetoğlu, A., Sezer, E., & Erol, S. (2021). Development of antimicrobial whey protein-based film containing silver nanoparticles biosynthesised by Aspergillus Niger. *International Journal of Food Science & Technology*, 56(2), 965–973. 10.1111/iifs.14749.
- Çakmak, H., Özselek, Y., Turan, O. Y., Fıratlıgil, E., & Karbancioğlu-Güler, F. (2020). Whey protein isolate edible films incorporated with essential oils: Antimicrobial activity and barrier properties. *Polymer Degradation and Stability*, 179, Article 109285. 10.1016/j.polymdegradstab.2020.109285.
- Cao, N., Fu, Y., & He, J. (2007). Preparation and physical properties of soy protein isolate and gelatin composite films. *Food Hydrocolloids*, 21(7), 1153–1162. 10.1016/j.foodhyd.2006.09.001.
- Cardador, M. J., & Gallego, M. (2016). Origin of haloacetic acids in milk and dairy products. Food Chemistry, 196, 750–756. 10.1016/j.foodchem.2015.10.011.
- Cárdenas, G., Díaz, J., Meléndrez, M. F., & Cruzat, C. (2008). Physicochemical properties of edible films from chitosan composites obtained by microwave heating. *Polymer Bulletin*, 61(6), 737–748. 10.1007/s00289-008-0994-7.
- Carpiné, D., Dagostin, J. L. A., Bertan, L. C., & Mafra, M. R. (2015). Development and Characterization of Soy Protein Isolate Emulsion-Based Edible Films with Added Coconut Oil for Olive Oil Packaging: Barrier, Mechanical, and Thermal Properties. *Food and Bioprocess Technology*, 8(8), 1811–1823. 10.1007/s11947-015-1538-4.
- Castro, F. V. R., Andrade, M. A., Sanches Silva, A., Fátima Vaz, M., Vilarinho, F., Doutor, R., Jorge, I. P., & Cruz, A. P. (2019). The Contribution of a Whey Protein Film Incorporated with Green Tea Extract to Minimize the Lipid Oxidation of Salmon (Salmo salar L.). Mdpi.Com. 10.3390/foods8080327.
- Cazón, P., Velazquez, G., Ramírez, J. A., & Vázquez, M. (2017). Polysaccharide-based films and coatings for food packaging: A review. *Food Hydrocolloids*, 68, 136–148. 10.1016/j.foodhyd.2016.09.009.
- Ceballos, R. L., Ochoa-Yepes, O., Goyanes, S., Bernal, C., & Famá, L. (2020). Effect of yerba mate extract on the performance of starch films obtained by extrusion and compression molding as active and smart packaging. *Carbohydrate Polymers*, 244, Article 116495. 10.1016/j.carbpol.2020.116495.
- Chavoshizadeh, S., Pirsa, S., & Mohtarami, F. (2020). Conducting/smart color film based on wheat gluten/chlorophyll/polypyrrole nanocomposite. *Food Packaging and Shelf Life*, 24, Article 100501 January. 10.1016/j.fpsl.2020.100501.
- Chen, Y., Ye, R., & Liu, J. (2014). Effects of different concentrations of ethanol and isopropanol on physicochemical properties of zein-based films. *Industrial Crops and Products*, 53, 140–147. 10.1016/j.indcrop.2013.12.034.

- Cheng, Y., Gao, S., Wang, W., Hou, H., & Lim, L.-T. (2022). Low temperature extrusion blown ε-polylysine hydrochloride-loaded starch/gelatin edible antimicrobial films. *Carbohydrate Polymers*, 278, 18990. 10.1016/j.carbpol.2021.118990.
- Cheng, S. Y., Wang, B. J., & Weng, Y. M. (2015). Antioxidant and antimicrobial edible zein/chitosan composite films fabricated by incorporation of phenolic compounds and dicarboxylic acids. *LWT - Food Science and Technology*, 63(1), 115–121. 10.1016/j.lwt.2015.03.030.
- Chevalier, E., Chaabani, A., Assezat, G., Prochazka, F., & Oulahal, N. (2018). Casein/wax blend extrusion for production of edible films as carriers of potassium sorbate—A comparative study of waxes and potassium sorbate effect. *Food Packaging and Shelf Life*, 16, 41–50. 10.1016/j.fpsl.2018.01.005.
- Chiralt, A., González-Martínez, C., Vargas, M., & Atarés, L. (2018). 18—Edible films and coatings from proteins. In R. Y. Yada (Ed.), *Proteins in Food Processing* (pp. 477–500). Woodhead Publishing. 10.1016/B978-0-08-100722-8.00019-X.
- Cho, S.-W., Gällstedt, M., Johansson, E., & Hedenqvist, M. S (2011). Injection-molded nanocomposites and materials based on wheat gluten. *International journal of biological* macromolecules, 48(1), 146–152. 10.1016/j.ijbiomac.2010.10.012.
- Chodijah, S., Husaini, A., & Zaman, M. (2019). Extraction of pectin from banana peels (musa paradiasica fomatypica) for biodegradable plastic films. *Journal of Physics: Conference Series (Vol. 1167, No. 1, p. 012061).* 10.1088/1742-6596/1167/1/012061.
- Coppola, D., Oliviero, M., Vitale, G. A., Lauritano, C., D'Ambra, I., Iannace, S., & de Pascale, D. (2020). Marine collagen from alternative and sustainable sources: Extraction, processing and applications. Marine Drugs (Vol. 18, Issue 4, p. 214). MDPI AG. 10.3390/md18040214.
- Cristine De Souza, K., Correa, L. G., Tamires, Vieira Da Silva, B., Fernandes, T., Moreira, M., Anielle De Oliveira, Sakanaka, L. S., Dias, M. I., Barros, L., Ferreira, I. C. F. R., Valderrama, P., Fernanda, &, Leimann, V., & Shirai, M. A (2020). Soy Protein Isolate Films Incorporated with Pinhão (Araucaria angustifolia (Bertol.) Kuntze) Extract for Potential Use as Edible Oil Active Packaging. *Food Bioprocess Technol, 13*, 998–1008. 10.1007/s11947-020-02454-5.
- Cruz-Diaz, K., Cobos, Á., Fernández-Valle, M. E., Díaz, O., & Cambero, M. I. (2019). Characterization of edible films from whey proteins treated with heat, ultrasounds and/or transglutaminase. Application in cheese slices packaging. *Food Packaging and Shelf Life*, 22, Article 100397. 10.1016/j.fpsl.2019.100397.
- da Rocha, M., Alemán, A., Romani, V. P., López-Caballero, M. E., Gómez-Guillén, M. C., Montero, P., & Prentice, C. (2018). Effects of agar films incorporated with fish protein hydrolysate or clove essential oil on flounder (*Paralichthys orbignyanus*) fillets shelflife. *Food Hydrocolloids*, 81, 351–363. 10.1016/j.foodhyd.2018.03.017.
- Dang, K. M., & Yoksan, R. (2015). Development of thermoplastic starch blown film by incorporating plasticized chitosan. *Carbohydrate Polymers*, 115, 575–581. 10.1016/j.carbpol.2014.09.005.
- Dash, K. K., Ali, N. A., Das, D., & Mohanta, D. (2019). Thorough evaluation of sweet potato starch and lemon-waste pectin based-edible films with nano-titania inclusions for food packaging applications. *International Journal of Biological Macromolecules*, 139, 449– 458. 10.1016/j.ijbiomac.2019.07.193.
- Debeaufort, F., Quezada-Gallo, J., & Voilley, A. (1998). Edible Films and Coatings : Tomorrow 's Packagings : A Review. Critical Reviews in Food Science, 38(4), 299–313. 10.1080/10408699891274219.
- Dehghani, S., Hosseini, S. V., & Regenstein, J. M. (2018). Edible films and coatings in seafood preservation: A review. In Food Chemistry: 240 (pp. 505–513). Elsevier Ltd. 10.1016/j.foodchem.2017.07.034.
- Deng, L., Li, X., Miao, K., Mao, X., Han, M., Li, D., Mu, C., & Ge, L. (2020). Development of Disulfide Bond Crosslinked Gelatin/*e*-Polylysine Active Edible Film with Antibacterial and Antioxidant Activities. *Food and Bioprocess Technology*, 13(4), 577–588. 10.1007/s11947-020-02420-1.
- Dhall, R. K. (2013). Advances in edible coatings for fresh fruits and vegetables: A review. Critical Reviews in Food Science and Nutrition, 53(5), 435–450. 10.1080/10408398.2010.541568.
- Dhanapal, A., Rajamani, L., & Banu, M. (2012). Edible films from Polysaccharides. Food Science and Quality Management, 3(1), 9–18.
- Dhumal, C. V., & Sarkar, P. (2018). Composite edible films and coatings from foodgrade biopolymers. Journal of Food Science and Technology, 55(11), 4369–4383. 10.1007/s13197-018-3402-9.
- Diab, T., Biliaderis, C. G., Gerasopoulos, D., & Sfakiotakis, E. (2001). Physicochemical properties and application of pullulan edible films and coatings in fruit preservation. *Journal of the Science of Food and Agriculture*, 81(10), 988–1000. 10.1002/jsfa.883.
- Di Giuseppe, F. A., Volpe, S., Cavella, S., Masi, P., & Torrieri, E. (2022). Physical properties of active biopolymer films based on chitosan, sodium caseinate, and rosemary essential oil. Food Packaging and Shelf Life, 32, Article 100817. 10.1016/j.fpsl.2022.100817.
- Dong, M., Tian, L., Li, J., Jia, J., Dong, Y., Tu, Y., Liu, X., Tan, C., & Duan, X. (2022). Improving physicochemical properties of edible wheat gluten protein films with proteins, polysaccharides and organic acid. *LWT*, 154, Article 112868. 10.1016/j.lwt.2021.112868.
- Ebrahimi, B., Mohammadi, R., Rouhi, M., Mortazavian, A. M., Shojaee-Aliabadi, S., & Koushki, M. R. (2018). Survival of probiotic bacteria in carboxymethyl cellulose-based edible film and assessment of quality parameters. *LWT*, *87*, 54–60. 10.1016/j.lwt.2017.08.066.
- Escamilla-García, M., Calderón-Domínguez, G., Chanona-Pérez, J. J., Farrera-Rebollo, R. R., Andraca-Adame, J. A., Arzate-Vázquez, I., Mendez-Mendez, J. V., & Moreno-Ruiz, L. A. (2013). Physical and structural characterisation of zein and chitosan edible films using nanotechnology tools. *International Journal of Biological Macromolecules*, 61, 196–203. 10.1016/j.ijbiomac.2013.06.051.
- Espitia, P. J. P., Batista, R. A., Azeredo, H. M. C., & Otoni, C. G. (2016). Probiotics and their potential applications in active edible films and coatings. *Food Research International*, 90, 42–52. 10.1016/j.foodres.2016.10.026.

- Espitia, P. J. P., & Otoni, C. G. (2018). Nanotechnology and edible films for food packaging applications. In *Bio-based Materials for Food Packaging: Green* and Sustainable Advanced Packaging Materials (pp. 125–145). Singapore: Springer. 10.1007/978-981-13-1909-9\_6.
- Fagundes, C., Palou, L., Monteiro, A. R., & Pérez-Gago, M. B. (2015). Hydroxypropyl methylcellulose-beeswax edible coatings formulated with antifungal food additives to reduce alternaria black spot and maintain postharvest quality of cold-stored cherry tomatoes. *Scientia Horticulturae*, 193, 249–257. 10.1016/j.scienta.2015.07.027.
- Fajardo, P., Martins, J. T., Fuciños, C., Pastrana, L., Teixeira, J. A., & Vicente, A. A. (2010). Evaluation of a chitosan-based edible film as carrier of natamycin to improve the storability of Saloio cheese. *Journal of Food Engineering*, 101(4), 349–356. 10.1016/j.jfoodeng.2010.06.029.
- Fakhouri, F. M., Martelli, S. M., Caon, T., Velasco, J. I., Buontempo, R. C., Bilck, A. P., & Innocentini Mei, L. H. (2017). The effect of fatty acids on the physicochemical properties of edible films composed of gelatin and gluten proteins. *LWT - Food Science* and Technology. 10.1016/j.lwt.2017.08.056.
- Fakhreddin Hosseini, S., Rezaei, M., Zandi, M., & Ghavi, F. F. (2013). Preparation and functional properties of fish gelatin-chitosan blend edible films. *Food Chemistry*, 136(3–4), 1490–1495. 10.1016/j.foodchem.2012.09.081.
- Falguera, V., Quintero, J. P., Jiménez, A., Muñoz, J. A., & Ibarz, A. (2011). Edible films and coatings: Structures, active functions and trends in their use. *Trends in Food Science* and Technology, 22(6), 292–303. 10.1016/j.tifs.2011.02.004.
- Fan, Y., Yang, J., Duan, A., & Li, X. (2021). Pectin/sodium alginate/xanthan gum edible composite films as the fresh-cut package. *International Journal of Biological Macromolecules*, 181, 1003–1009. 10.1016/j.ijbiomac.2021.04.111.
- Fangfang, Z., Xinpeng, B., Wei, G., Wang, G., Shi, Z., & Jun, C. (2020). Effects of virgin coconut oil on the physicochemical, morphological and antibacterial properties of potato starch-based biodegradable films. *International Journal of Food Science and Technology*, 55(1), 192–200. 10.1111/ijfs.14262.
- Farajzadeh, F., Motamedzadegan, A., Shahidi, S.-A., & Hamzeh, S. (2016). The effect of chitosan-gelatin coating on the quality of shrimp (*Litopenaeus vannamei*) under refrigerated condition. *Food control*, 67, 163–170. 10.1016/j.foodcont.2016.02.040.
- Fernandes, L. M., Guimarães, J. T., Pimentel, T. C., Esmerino, E. A., Freitas, M. Q., Carvalho, C. W. P., Cruz, A. G., & Silva, M. C. (2020). Chapter 7—Edible whey protein films and coatings added with prebiotic ingredients. In F. J. Barba, P. Putnik, & D. B. Kovačević (Eds.), Agri-Food Industry Strategies for Healthy Diets and Sustainability (pp. 177–193). Academic Press. 10.1016/B978-0-12-817226-1.00007-2.
- Flores-López, M. L., Cerqueira, M. A., de Rodríguez, D. J., & Vicente, A. A. (2016). Perspectives on Utilization of Edible Coatings and Nano-laminate Coatings for Extension of Postharvest Storage of Fruits and Vegetables. *Food Engineering Reviews*, 8(3), 292–305. 10.1007/s12393-015-9135-x.
- Friesen, K., Chang, C., & Nickerson, M. (2015). Incorporation of phenolic compounds, rutin and epicatechin, into soy protein isolate films: Mechanical, barrier and cross-linking properties. *Food Chemistry*, 172, 18–23. 10.1016/j.foodchem.2014.08.128.
- Future, M. R. (2021). Edible Packaging Market worth USD 2.14 billion by 2030, registering a CAGR of 6.79%—Report by Market Research Future (MRFR). *GlobeNewswire News Room* https://www. globenewswire.com/news-release/2021/12/15/2353020/0/en/Edible-Packaging-Market-worth-USD-2-14-billion-by-2030-registering-a-CAGR-of-6-79-Report-by-Market-Research-Future-MRFR.html accessed on 18/03/2022.
- Galus, S., & Kadzińska, J. (2015). Food applications of emulsion-based edible films and coatings. Trends in Food Science & Technology, 45(2), 273–283. 10.1016/j.tifs.2015.07.011.
- Galus, S., & Kadzińska, J. (2016). Whey protein edible films modified with almond and walnut oils. Food Hydrocolloids, 52, 78–86. 10.1016/j.foodhyd.2015.06.013.
- Gao, Z. hua, Zhang, Y. hong, Fang, B., Zhang, L. peng, & Shi, J. (2015). The effects of thermal-acid treatment and crosslinking on the water resistance of soybean protein. *Industrial Crops and Products*, 74, 122–131. 10.1016/j.indcrop.2015.04.026.
- Ge, L., Zhu, M., Xu, Y., Li, X., Li, D., & Mu, C. (2017). Development of Antimicrobial and Controlled Biodegradable Gelatin-Based Edible Films Containing Nisin and Amino-Functionalized Montmorillonite. *Food and Bioprocess Technology*, 10(9), 1727–1736. 10.1007/s11947-017-1941-0.
- Genevois, C. E., de Escalada Pla, M. F., & Flores, S. K. (2016). Application of edible coatings to improve global quality of fortified pumpkin. *Innovative Food Science & Emerging Technologies*, 33, 506–514. 10.1016/j.ifset.2015.11.001.
- Ghasemlou, M., Khodaiyan, F., & Oromiehie, A. (2011). Rheological and structural characterisation of film-forming solutions and biodegradable edible film made from kefiran as affected by various plasticizer types. *International Journal of Biological Macromolecules*, 49(4), 814–821. 10.1016/j.ijbiomac.2011.07.018.
- Gheorghita (Puscaselu), R., Amariei, S., Norocel, L., & Gutt, G. (2020). New Edible Packaging Material with Function in Shelf Life Extension: Applications for the Meat and Cheese Industries. *Foods*, 9(5), 562. 10.3390/foods9050562.
- Giteru, S. G., Cridge, B., Oey, I., Ali, A., & Altermann, E. (2020). In-vitro degradation and toxicological assessment of pulsed electric fields crosslinked zein-chitosanpoly(vinyl alcohol) biopolymeric films. *Food and Chemical Toxicology*, 135, Article 111048. 10.1016/j.fct.2019.111048.
- Gómez-Estaca, J., López de Lacey, A., López-Caballero, M. E., Gómez-Guillén, M. C., & Montero, P. (2010). Biodegradable gelatin-chitosan films incorporated with essential oils as antimicrobial agents for fish preservation. *Food Microbiology*, 27(7), 889–896. 10.1016/j.fm.2010.05.012.
- Gorrasi, G., & Bugatti, V. (2016). Edible bio-nano-hybrid coatings for food protection based on pectins and LDH-salicylate: Preparation and analysis of physical properties. *LWT - Food Science and Technology*, 69, 139–145. 10.1016/j.lwt.2016.01.038.
- Groh, K. J., Backhaus, T., Carney-Almroth, B., Geueke, B., Inostroza, P. A., Lennquist, A., Leslie, H. A., Maffini, M., Slunge, D., Trasande, L., Warhurst, A. M., & Muncke, J. (2019). Overview of known plastic packaging-associated chem-

#### A.K. V, M. Hasan, S. Mangaraj et al.

icals and their hazards. Science of the Total Environment, 651, 3253-3268. 10.1016/j.scitotenv.2018.10.015.

- Günlü, A., & Koyun, E. (2013). Effects of Vacuum Packaging and Wrapping with Chitosan-Based Edible Film on the Extension of the Shelf Life of Sea Bass (*Dicentrarchus labrax*) Fillets in Cold Storage (4°C). Food and Bioprocess Technology, 6(7), 1713–1719. 10.1007/s11947-012-0833-6.
- Han, K., Liu, Y., Liu, Y., Huang, X., & Sheng, L. (2020). Characterization and film-forming mechanism of egg white/pullulan blend film. *Food Chemistry*, 315, Article 126201. 10.1016/j.foodchem.2020.126201.
- Hasan, M., V, A. K., Maheshwari, C., & Mangraj, S (2020). Biodegradable and edible film: A counter to plastic pollution. *International Journal of Chemical Studies*, 8(1), 2242– 2245. 10.22271/chemi.2020.v8.i1ah.8606.
- Hashemi, S. M. B., & Mousavi Khaneghah, A. (2017). Characterization of novel basilseed gum active edible films and coatings containing oregano essential oil. *Progress in Organic Coatings*, 110, 35–41 April. 10.1016/j.porgcoat.2017.04.041.
- Hassan, B., Chatha, S. A. S., Hussain, A. I., Zia, K. M., & Akhtar, N. (2018). Recent advances on polysaccharides, lipids and protein based edible films and coatings: A review. *International Journal of Biological Macromolecules*, 109, 1095–1107. 10.1016/J.IJBIOMAC.2017.11.097.
- He, J., Wang, R., Feng, W., Chen, Z., & Wang, T. (2020). Design of novel edible hydrocolloids by structural interplays between wheat gluten proteins and soy protein isolates. *Food Hydrocolloids*, 100, Article 105395. 10.1016/j.foodhyd.2019.105395.
- Hemsri, S., Asandei, A. D., Grieco, K., & Parnas, R. S. (2011). Biopolymer composites of wheat gluten with silica and alumina. *Composites Part A: Applied Science and Manufacturing*, 42(11), 1764–1773. 10.1016/j.compositesa.2011.07.032.
- Hernandez-Izquierdo, V. M., & Krochta, J. M. (2008). Thermoplastic processing of proteins for film formation - A review. *Journal of Food Science*, 73(2), 30–39. 10.1111/j.1750-3841.2007.00636.x.
- Hromiš, N., Lazić, V., Popović, S., Šuput, D., Bulut, S., Kravić, S., & Romanić, R. (2022). The possible application of edible pumpkin oil cake film as pouches for flaxseed oil protection. *Food Chemistry*, 371, Article 131197. 10.1016/j.foodchem.2021.131197.
- Huang, X., Luo, X., Liu, L., Dong, K., Yang, R., Lin, C., Song, H., Li, S., & Huang, Q. (2020). Formation mechanism of egg white protein/κ-Carrageenan composite film and its application to oil packaging. *Food Hydrocolloids*, 105(February), Article 105780. 10.1016/j.foodhyd.2020.105780.
- Huntrakul, K., Yoksan, R., Sane, A., & Harnkarnsujarit, N. (2020). Effects of pea protein on properties of cassava starch edible films produced by blown-film extrusion for oil packaging. *Food Packaging and Shelf Life, 24*, Article 100480 February. 10.1016/j.fpsl.2020.100480.
- Imre, B., García, L., Puglia, D., & Vilaplana, F. (2019). Reactive compatibilization of plant polysaccharides and biobased polymers: Review on current strategies, expectations and reality. *Carbohydrate Polymers*, 209, 20–37. 10.1016/j.carbpol.2018.12.082.
- Kowalczyk, D. (2011). Effect of edible protein-wax coating on the post-harvest stability of Brussels sprouts stored in simulated conditions of trade. *Food Science Technology Quality*, 18(6).
- Janjarasskul, T., & Krochta, J. M. (2010). Edible Packaging Materials. Annual Review of Food Science and Technology, 1(1), 415–448. 10.1146/annurev.food.080708.100836.
- Jebalia, I., Maigret, J. E., Réguerre, A. L., Novales, B., Guessasma, S., Lourdin, D., Della Valle, G., & Kristiawan, M. (2019). Morphology and mechanical behaviour of peabased starch-protein composites obtained by extrusion. *Carbohydrate Polymers, 223*, Article 115086. 10.1016/j.carbpol.2019.115086.
- Jeevahan, J. J., Chandrasekaran, M., Venkatesan, S. P., Sriram, V., Joseph, G. B., Mageshwaran, G., & Durairaj, R. B. (2020). Scaling up difficulties and commercial aspects of edible films for food packaging: A review. *Trends in Food Science & Technology*, 100, 210–222. 10.1016/j.tifs.2020.04.014.
- Jouki, M., Khazaei, N., Ghasemlou, M., & Hadinezhad, M. (2013). Effect of glycerol concentration on edible film production from cress seed carbohydrate gum. *Carbohydrate Polymers*, 96(1), 39–46. 10.1016/j.carbpol.2013.03.077.
- Jridi, M., Boughriba, S., Abdelhedi, O., Nciri, H., Nasri, R., Kchaou, H., Kaya, M., Sebai, H., Zouari, N., & Nasri, M. (2019). Investigation of physicochemical and antioxidant properties of gelatin edible film mixed with blood orange (*Citrus sinensis*) peel extract. *Food Packaging and Shelf Life*, 21, Article 100342. 10.1016/j.fpsl.2019.100342.
- Kaewprachu, P., Osako, K., Benjakul, S., & Rawdkuen, S. (2016). Effect of protein concentrations on the properties of fish myofibrillar protein based film compared with PVC film. Journal of Food Science and Technology, 53(4), 2083–2091. 10.1007/s13197-016-2170-7.
- Kakaei, S., & Shahbazi, Y. (2016). Effect of chitosan-gelatin film incorporated with ethanolic red grape seed extract and Ziziphora clinopodioides essential oil on survival of Listeria monocytogenes and chemical, microbial and sensory properties of minced trout fillet. LWT-Food Science and Technology, 72, 432–438. 10.1016/j.lwt.2016.05.021.
- Kaya, S., & Kaya, A. (2000). Microwave drying effects on properties of whey protein isolate edible films. *Journal of Food Engineering*, 43(2), 91–96. 10.1016/S0260-8774(99)00136-3.
- Khaoula, K., Cristina, P., Sylvie, B., Sthane, D., Jo, H., Khwaldia, K., Perez, C., Banon, S., Desobry, S., & Hardy, J. (2004). Milk Proteins for Edible Films and Coatings. *Critical Reviews in Food Science and Nutrition*, 44(4), 239–251. 10.1080/10408690490464906.
- Khazaei, N., Esmaiili, M., & Emam-Djomeh, Z. (2016). Effect of active edible coatings made by basil seed gum and thymol on oil uptake and oxidation in shrimp during deep-fat frying. *Carbohydrate polymers*, 137, 249–254. 10.1016/j.carbpol.2015.10.084.
- Kim, D., & Min, S. C. (2012). Trout Skin Gelatin-Based Edible Film Development. Journal of Food Science, 77(9), E240–E246. 10.1111/j.1750-3841.2012.02880.x.
- Krishna, M., Nindo, C. I., & Min, S. C. (2012). Development of fish gelatin edible films using extrusion and compression molding. *Journal of Food Engineering*, 108(2), 337– 344. 10.1016/j.jfoodeng.2011.08.002.

- Kumar, S., Mukherjee, A., & Dutta, J. (2020). Chitosan based nanocomposite films and coatings: Emerging antimicrobial food packaging alternatives. *Trends in Food Science* and Technology, 97, 196–209. 10.1016/j.tifs.2020.01.002.
- Kumari, M., Mahajan, H., Joshi, R., & Gupta, M. (2017). Development and structural characterization of edible films for improving fruit quality. *Food Packaging and Shelf Life*. 10.1016/j.fpsl.2017.02.003.
- Lalnunthari, C., Devi, L. M., & Badwaik, L. S. (2020). Extraction of protein and pectin from pumpkin industry by-products and their utilization for developing edible film. *Journal* of Food Science and Technology, 57(5), 1807–1816. 10.1007/s13197-019-04214-6.
- Lee, J. Y., Park, H. J., Lee, C. Y., & Choi, W. Y. (2003). Extending shelf-life of minimally processed apples with edible coatings and antibrowning agents. *LWT - Food Science* and Technology, 36(3), 323–329. 10.1016/S0023-6438(03)00014-8.
- Li, Y., Cao, C., Pei, Y., Liu, X., & Tang, K. (2019). Preparation and properties of microfibrillated chitin/gelatin composites. *International Journal of Biological Macromolecules*, 130, 715–719. 10.1016/j.ijbiomac.2019.03.014.
- Licciardello, F., Kharchoufi, S., Muratore, G., & Restuccia, C. (2018). Effect of edible coating combined with pomegranate peel extract on the quality maintenance of white shrimps (*Parapenaeus longirostris*) during refrigerated storage. *Food packaging and shelf life*, 17, 114–119. 10.1016/j.fpsl.2018.06.009.
- Lin, D., Zheng, Y., Huang, Y., Ni, L., Zhao, J., Huang, C., Chen, X., Chen, X., Wu, Z., Wu, D., Chen, H., Zhang, Q., Qin, W., & Xing, B. (2020). Investigation of the structural, physical properties, antioxidant, and antimicrobial activity of chitosan- nano-silicon aerogel composite edible films incorporated with okara powder. *Carbohydrate Polymers*, 250, Article 116842. 10.1016/j.carbpol.2020.116842.
- Lin, H. C., Wang, B. J., & Weng, Y. M. (2020). Development and characterization of sodium caseinate edible films cross-linked with genipin. LWT - Food Science and Technology Journal, 118(June), Article 108813. 10.1016/j.lwt.2019.108813.
- Lisitsyn, A., Semenova, A., Nasonova, V., Polishchuk, E., Revutskaya, N., Kozyrev, I., & Kotenkova, E. (2021). Approaches in Animal Proteins and Natural Polysaccharides Application for Food Packaging: Edible Film Production and Quality Estimation. *Polymers*, 13(10), 1592. 10.3390/polym13101592.
- Lopez, D., Marquez, A., Gutierrez-Cutino, M., Venegas-Yazigi, D., Bustos, R., & Matiacevich, S. (2017). Edible film with antioxidant capacity based on salmon gelatin and boldine. LWT - Food Science and Technology, 77, 160–169. 10.1016/j.lwt.2016.11.039.
- López de Lacey, A. M., López-Caballero, M. E., & Montero, P. (2014). Agar films containing green tea extract and probiotic bacteria for extending fish shelf-life. *LWT - Food Science* and Technology, 55(2), 559–564. 10.1016/j.lwt.2013.09.028.
- Madsen, M., Westh, P., Khan, S., Ipsen, R., Almdal, K., Aachmann, F. L., & Svensson, B. (2021). Impact of Alginate Mannuronic-Guluronic Acid Contents and pH on Protein Binding Capacity and Complex Size. *Biomacromolecules*. 10.1021/acs.biomac.0c01485.
- Mahcene, Z., Khelil, A., Hasni, S., Bozkurt, F., Goudjil, M. B., & Tornuk, F. (2020). Homemade cheese preservation using sodium alginate based on edible film incorporating essential oils. *Journal of Food Science and Technology*. 10.1007/s13197-020-04753-3.
- Malakar, A., Kanel, S. R., Ray, C., Snow, D. D., & Nadagouda, M. N. (2021). Nanomaterials in the environment, human exposure pathway, and health effects: A review. *Science* of the Total Environment, 759, Article 143470. 10.1016/j.scitotenv.2020.143470.
- Malik, G. K., & Mitra, J. (2021). Zinc oxide nanoparticle synthesis, characterization, and their effect on mechanical, barrier, and optical properties of hpmc-based edible film. *Food and Bioprocess Technology*, 14(3), 441–456. 10.1007/s11947-020-02566-y.
- Martins, J. T., Cerqueira, M. A., Souza, B. W. S., Carmo Avides, M. D. O., & Vicente, A. A. (2010). Shelf life extension of ricotta cheese using coatings of galactomannans from nonconventional sources incorporating nisin against listeria monocytogenes. *Journal of Agricultural and Food Chemistry*, 58(3), 1884–1891. 10.1021/if902774z.
- Mehyar, G. F., Al-Ismail, K., Han, J. H., & Chee, G. W. (2012). Characterization of Edible Coatings Consisting of Pea Starch, Whey Protein Isolate, and Carnauba Wax and their Effects on Oil Rancidity and Sensory Properties of Walnuts and Pine Nuts. *Wiley Online Library*, 77(2). 10.1111/j.1750-3841.2011.02559.x.
- Mellinas, C., Valdés, A., Ramos, M., Burgos, N., Garrigós, M. del C., & Jiménez, A (2016). Active edible films: Current state and future trends. *Journal of Applied Polymer Science*, (2), 133. 10.1002/app.42631.
- Moghadam, M., Salami, M., Mohammadian, M., Khodadadi, M., & Emam-Djomeh, Z. (2020). Development of antioxidant edible films based on mung bean protein enriched with pomegranate peel. *Food Hydrocolloids*, 104, Article 105735. 10.1016/j.foodhyd.2020.105735.
- Mohamed, S. A. A., El-Sakhawy, M., & El-Sakhawy, M. A. M. (2020). Polysaccharides, Protein and Lipid -Based Natural Edible Films in Food Packaging: A Review. Carbohydrate Polymers, 238, Article 116178. 10.1016/j.carbpol.2020.116178.
- Moradi, M., Tajik, H., Razavi Rohani, S. M., & Mahmoudian, A. (2016). Antioxidant and antimicrobial effects of zein edible film impregnated with Zataria multiflora Boiss. essential oil and monolaurin. LWT - Food Science and Technology, 72, 37–43. 10.1016/j.lwt.2016.04.026.
- Moradi, M., Guimarães, J. T., & Sahin, S. (2021). Current applications of exopolysaccharides from lactic acid bacteria in the development of food active edible packaging. *Current Opinion in Food Science*, 40, 33–39. 10.1016/j.cofs.2020.06.001.
- Morales-Contreras, B. E., Wicker, L., Rosas-Flores, W., Contreras-Esquivel, J. C., Gallegos-Infante, J. A., Reyes-Jaquez, D., & Morales-Castro, J. (2020). Apple pomace from variety "Blanca de Asturias" as sustainable source of pectin: Composition, rheological, and thermal properties. *LWT*, 117, Article 108641. 10.1016/j.lwt.2019.108641.
- Murrieta-Martínez, C. L., Soto-Valdez, H., Pacheco-Aguilar, R., Torres-Arreola, W., Rodríguez-Felix, F., & Márquez Ríos, E. (2018). Edible protein films: Sources and behavior. *Packaging Technology and Science*, 31(3), 113–122. 10.1002/pts.2360.

- Nair, M. S., Tomar, M., Punia, S., Kukula-Koch, W., & Kumar, M. (2020). Enhancing the functionality of chitosan-and alginate-based active edible coatings/films for the preservation of fruits and vegetables: A review. *International Journal of Biological Macromolecules*, 164, 304–320. 10.1016/j.ijbiomac.2020.07.083.
- Nandane, A. S., & Jain, R. K. (2018). Optimization of Formulation and Process Parameters for Soy Protein-Based Edible Film Using Response Surface Methodology. *Journal of Packaging Technology and Research*, 2(3), 203–210. 10.1007/s41783-018-0045-2.
- Naseri, H. R., Beigmohammadi, F., Mohammadi, R., & Sadeghi, E. (2020). Production and characterization of edible film based on gelatin–chitosan containing *Ferulago angulate* essential oil and its application in the prolongation of the shelf life of turkey meat. *Journal of Food Processing and Preservation*, 44(8), e14558. 10.1111/jfpp.14558.
- Ngatirah, N., Ruswanto, A., & Sunardi, S. (2022). Effect of Hydroxypropyl methylcellulose from oil palm empty fruit bunch on oil uptake and physical properties of French fries. *Food Science and Technology*, 42. 10.1590/fst.110421.
- Navarro-Tarazaga, M. L., Massa, A., & Pérez-Gago, M. B. (2011). Effect of beeswax content on hydroxypropyl methylcellulose-based edible film properties and postharvest quality of coated plums (Cv. Angeleno). LWT - Food Science and Technology, 44(10), 2328–2334. 10.1016/j.lwt.2011.03.011.
- Nguyen, T. T., Thi Dao, U. T., Thi Bui, Q. P., Bach, G. L., Ha Thuc, C. N., & Ha Thuc, H. (2020). Enhanced antimicrobial activities and physiochemical properties of edible film based on chitosan incorporated with *Sonneratia caseolaris* (L.) Engl. leaf extract. *Progress in Organic Coatings*, 140, Article 105487. 10.1016/j.porgcoat.2019.105487.
- Nunes, J. C., Melo, P. T. S., Lorevice, M. V., Aouada, F. A., & de Moura, M. R. (2020). Effect of green tea extract on gelatin-based films incorporated with lemon essential oil. Journal of Food Science and Technology, 58(1), 1–8. 10.1007/s13197-020-04469-4.
- Nussinovitch, A. (2013). Biopolymer Films and Composite Coatings. In Handbook of Biopolymers and Biodegradable Plastics (pp. 295–327). 10.1016/B978-1-4557-2834-3.00013-6.
- Ochoa-Yepes, O., Di Giogio, L., Goyanes, S., Mauri, A., & Famá, L. (2019). Influence of process (extrusion/thermo-compression, casting) and lentil protein content on physicochemical properties of starch films. *Carbohydrate Polymers*, 208, 221–231. 10.1016/j.carbpol.2018.12.030.
- Ochoa, E., Saucedo-Pompa, S., Rojas-Molina, R., Garza, H. D. L., Charles-Rodríguez, A. V., & Aguilar, C. N. (2011). Evaluation of a candelilla wax-based edible coating to prolong the shelf-life quality and safety of apples. *American journal of agricultural and biological* sciences, 6(1), 92–98. 10.3844/ajabssp.2011.92.98.
- Ochoa, T. A., García-Almendárez, B. E., Reyes, A. A., Pastrana, D. M. R., López, G. F. G., Belloso, O. M., & González, C. R. (2017). Design and Characterization of Corn Starch Edible Films Including Beeswax and Natural Antimicrobials. *Food and Bioprocess Technology*, 10(1), 103–114. 10.1007/s11947-016-1800-4.
- Oğuzhan Yıldız, P., & Yangılar, F. (2016). Effects of Different Whey Protein Concentrate Coating on Selected Properties of Rainbow Trout (Oncorhynchusï¿mykiss) During Cold Storage (4°C). International Journal of Food Properties, 19(9), 2007–2015. 10.1080/10942912.2015.1092160.
- Omar-Aziz, M., Khodaiyan, F., Yarmand, M. S., Mousavi, M., Gharaghani, M., Kennedy, J. F., & Hosseini, S. S. (2021). Combined effects of octenylsuccination and beeswax on pullulan films: Water-resistant and mechanical properties. *Carbohydrate Polymers*, 255, Article 117471. 10.1016/j.carbpol.2020.117471.
- Oregel-Zamudio, E., Angoa-Pérez, M. V., Oyoque-Salcedo, G., Aguilar-González, C. N., & Mena-Violante, H. G. (2017). Effect of candelilla wax edible coatings combined with biocontrol bacteria on strawberry quality during the shelf-life. *Scientia Horticulturae*, 214, 273–279. 10.1016/j.scienta.2016.11.038.
- Orozco-Parra, J., Mejía, C. M., & Villa, C. C. (2020). Development of a bioactive synbiotic edible film based on cassava starch, inulin, and Lactobacillus casei. *Food Hydrocolloids*, 104, Article 105754. 10.1016/j.foodhyd.2020.105754.
- Otoni, C. G., Avena-Bustillos, R. J., Azeredo, H. M. C., Lorevice, M. V., Moura, M. R., Mattoso, L. H. C., & McHugh, T. H. (2017). Recent Advances on Edible Films Based on Fruits and Vegetables—A Review. *Comprehensive Reviews in Food Science and Food Safety*, 16(5), 1151–1169. 10.1111/1541-4337.12281.
- Otoni, C. G., Avena-Bustillos, R. J., Olsen, C. W., Bilbao-Sáinz, C., & McHugh, T. H. (2016). Mechanical and water barrier properties of isolated soy protein composite edible films as affected by carvacrol and cinnamaldehyde micro and nanoemulsions. *Food Hydrocolloids*, 57, 72–79. 10.1016/j.foodhyd.2016.01.012.
- Pal, N., Dubey, P., Gopinath, P., & Pal, K. (2017). Combined effect of cellulose nanocrystal and reduced graphene oxide into poly-lactic acid matrix nanocomposite as a scaffold and its anti-bacterial activity. *International Journal of Biological Macromolecules*, 95, 94–105. 10.1016/j.ijbiomac.2016.11.041.
- Peng, L., Wang, H., Dai, H., Fu, Y., Ma, L., Zhu, H., Yu, Y., Li, L., Wang, Q., & Zhang, Y. (2021). Preparation and characterization of gelatin films by transglutaminase cross-linking combined with ethanol precipitation or Hofmeister effect. *Food Hydrocolloids*, 113, Article 106421. 10.1016/j.foodhyd.2020.106421.
- Perdones, A., Escriche, I., Chiralt, A., & Vargas, M. (2016). Effect of chitosan-lemon essential oil coatings on volatile profile of strawberries during storage. *Food Chemistry*, 197, 979–986. 10.1016/j.foodchem.2015.11.054.
- Perez, V., Felix, M., Romero, A., & Guerrero, A. (2016). Characterization of pea proteinbased bioplastics processed by injection moulding. *Food and Bioproducts Processing*, 97, 100–108. 10.1016/j.fbp.2015.12.004.
- Pires, J. R. A., de Souza, V. G. L., & Fernando, A. L. (2018). Chitosan/montmorillonite bionanocomposites incorporated with rosemary and ginger essential oil as packaging for fresh poultry meat. *Food Packaging and Shelf Life*, 17, 142–149 June. 10.1016/j.fpsl.2018.06.011.
- Pizato, S., Chevalier, R. C., Dos Santos, M. F., Da Costa, T. S., Arévalo Pinedo, R., & Cortez Vega, W. R. (2019). Evaluation of the shelf-life extension of fresh-cut pineapple (*Smooth cayenne*) by application of different edible coatings. *British Food Journal*, 121(7), 1592–1604. 10.1108/BFJ-11-2018-0780.

- Poverenov, E., Zaitsev, Y., Arnon, H., Granit, R., Alkalai-Tuvia, S., Perzelan, Y., & Fallik, E. (2014). Effects of a composite chitosan–gelatin edible coating on postharvest quality and storability of red bell peppers. *Postharvest Biology and Technology*, 96, 106–109. 10.1016/j.postharvbio.2014.05.015.
- Puscaselu, R., Gutt, G., & Amariei, S. (2019). Rethinking the Future of Food Packaging: Biobased Edible Films for Powdered Food and Drinks. *Molecules*, 24(17), 3136. 10.3390/molecules24173136.
- Qiu, Y.-T., Wang, B.-J., & Weng, Y.-M. (2020). Preparation and characterization of genipin cross-linked and lysozyme incorporated antimicrobial sodium caseinate edible films. *Food Packaging and Shelf Life, 26*, Article 100601. 10.1016/j.fpsl.2020.100601.
- Ramos, Ó. L., Fernandes, J. C., Silva, S. I., Pintado, M. E., & Malcata, F. X. (2012). Edible Films and Coatings from Whey Proteins: A Review on Formulation, and on Mechanical and Bioactive Properties. *Critical Reviews in Food Science and Nutrition*, 52(6), 533–552. 10.1080/10408398.2010.500528.
- Remya, S., Mohan, C. O., Bindu, J., Sivaraman, G. K., Venkateshwarlu, G., & Ravishankar, C. N. (2016). Effect of chitosan based active packaging film on the keeping quality of chilled stored barracuda fish. *Journal of Food Science and Technology*, 53(1), 685–693. 10.1007/s13197-015-2018-6.
- Reyes-Avalos, M. C., Femenia, A., Minjares-Fuentes, R., Contreras-Esquivel, J. C., Aguilar-González, C. N., Esparza-Rivera, J. R., & Meza-Velázquez, J. A. (2016). Improvement of the quality and the shelf life of figs (*Ficus carica*) using an alginate-chitosan edible film. *Food and Bioprocess Technology*, 9(12), 2114–2124. 10.1007/s11947-016-1796-9.
- Rezaei, F., & Shahbazi, Y. (2018). Shelf-life extension and quality attributes of sauced silver carp fillet: A comparison among direct addition, edible coating and biodegradable film. LWT Food Science and Technology, 87, 122–133. 10.1016/j.lwt.2017.08.068.
- Rhim, J. W., & Shellhammer, T. H. (2005). 21—Lipid-based edible films and coatings. Innovations in Food Packaging, 362–383. 10.1016/B978-012311632-1/50053-X.
- Ribeiro, A. M., Estevinho, B. N., & Rocha, F. (2020). Preparation and Incorporation of Functional Ingredients in Edible Films and Coatings. *Food and Bioprocess Technology*, 14(2), 209–231. 10.1007/s11947-020-02528-4.
- Rodrigues, D. C., Cunha, A. P., Brito, E. S., Azeredo, H. M. C., & Gallão, M. I. (2016). Mesquite seed gum and palm fruit oil emulsion edible films: Influence of oil content and sonication. *Food Hydrocolloids*, 56, 227–235. 10.1016/j.foodhyd.2015.12.018.
- Rodriguez-Turienzo, L., Cobos, A., & Diaz, O. (2012). Effects of edible coatings based on ultrasound-treated whey proteins in quality attributes of frozen Atlantic salmon (Salmo salar). 10.1016/j.ifset.2011.12.003.
- Rodríguez, G. M., Sibaja, J. C., Espitia, P. J. P., & Otoni, C. G. (2020). Antioxidant active packaging based on papaya edible films incorporated with *Moringa oleifera* and ascorbic acid for food preservation. *Food Hydrocolloids*, 103, Article 105630 November 2019. 10.1016/j.foodhyd.2019.105630.
- Rossi Marquez, G., Di Pierro, P., Mariniello, L., Esposito, M., Giosafatto, C. V. L., & Porta, R. (2017). Fresh-cut fruit and vegetable coatings by transglutaminasecrosslinked whey protein/pectin edible films. *LWT - Food Science and Technology*, 75, 124–130. 10.1016/j.lwt.2016.08.017.
- Roşu, M.-C., Páll, E., Socaci, C., Măgeruşan, L., Pogăcean, F., Coroş, M., Turza, A., & Pruneanu, S. (2017). Cytotoxicity of methylcellulose-based films containing graphenes and curcumin on human lung fibroblasts. *Process Biochemistry*, 52, 243– 249. 10.1016/j.procbio.2016.10.002.
- Saberi, B., Thakur, R., Vuong, Q. V., Chockchaisawasdee, S., Golding, J. B., Scarlett, C. J., & Stathopoulos, C. E. (2016). Optimization of physical and optical properties of biodegradable edible films based on pea starch and guar gum. *Industrial Crops and Products*, 86, 342–352. 10.1016/j.indcrop.2016.04.015.
- Sáez, M. I., Suárez, M. D., & Martínez, T. F. (2020). Effects of alginate coating enriched with tannins on shelf life of cultured rainbow trout (*Oncorhynchus mykiss*) fillets. *LWT* - Food Science and Technology, 118, Article 108767.
- Sandhu, K. S., Sharma, L., Kaur, M., & Kaur, R. (2020). Physical, structural and thermal properties of composite edible films prepared from pearl millet starch and carrageenan gum: Process optimization using response surface methodology. *International Journal* of Biological Macromolecules, 143, 704–713. 10.1016/j.ijbiomac.2019.09.111.
- Senturk Parreidt, T., Müller, K., & Schmid, M. (2018). Alginate-Based Edible Films and Coatings for Food Packaging Applications. *Foods*, 7(10), 170. 10.3390/foods7100170.
- Seydim, A. C., Sarikus-Tutal, G., & Sogut, E. (2020). Effect of whey protein edible films containing plant essential oils on microbial inactivation of sliced Kasar cheese. Food Packaging and Shelf Life, 26, Article 100567. 10.1016/j.fpsl.2020.100567.
- Seyfzadeh, M., Motalebi, A. A., Kakoolaki, S., & Gholipour, H. (2013). Chemical, microbiological and sensory evaluation of gutted kilka coated with whey protein based edible film incorporated with sodium alginate during frozen storage. *Iranian Journal* of Fisheries Sciences, 12(1), 140–153.
- Sharma, L., & Singh, C. (2016). Sesame protein based edible films: Development and characterization. Food Hydrocolloids, 61, 139–147. 10.1016/j.foodhyd.2016.05.007.
- Shekarabi, A. S., Oromiehie, A. R., Vaziri, A., Ardjmand, M., & Safekordi, A. A. (2014). Investigation of the effect of nanoclay on the properties of quince seed mucilage edible films. *Food Science & Nutrition*, 2(6), 821–827. 10.1002/fsn3.177.
- Shen, Y., Ni, Z. J., Thakur, K., Zhang, J. G., Hu, F., & Wei, Z. J. (2021). Preparation and characterization of clove essential oil loaded nanoemulsion and pickering emulsion activated pullulan-gelatin based edible film. *International Journal of Biological Macromolecules*, 181, 528–539. 10.1016/j.ijbiomac.2021.03.133.
- Shroti, G. K., & Saini, C. S. (2022). Development of edible films from protein of brewer's spent grain: Effect of pH and protein concentration on physical, mechanical and barrier properties of films. Applied Food Research, 100043. 10.1016/j.afres.2022.100043.
- Soares, N. M. F., Oliveira, M. S. G., & Vicente, A. (2015). Effects of glazing and chitosanbased coating application on frozen salmon preservation during six-month storage in industrial freezing chambers. *LWT - Food Science and Technology*, 72, 285–291. 10.1016/j.lwt.2014.12.009.

- Soazo, M., P, L. M., Piccirilli, G. N., Delorenzi, N. J., & Verdini, R. A (2016). Antimicrobial and physicochemical characterization of whey protein concentrate edible films incorporated with liquid smoke. LWT - Food Science and Technology, 61, 524–531. 10.1016/j.lwt.2016.04.027.
- Soni, A., Gurunathan, K., Mendiratta, S. K., Talukder, S., Jaiswal, R. K., & Sharma, H. (2018). Effect of essential oils incorporated edible film on quality and storage stability of chicken patties at refrigeration temperature ( $4 \pm 1$  °C). Journal of Food Science and Technology, 55(9), 3538–3546. 10.1007/s13197-018-3279-7.
- Soukoulis, C., Behboudi-Jobbehdar, S., Macnaughtan, W., Parmenter, C., & Fisk, I. D. (2017). Stability of Lactobacillus rhamnosus GG incorporated in edible films: Impact of anionic biopolymers and whey protein concentrate. *Food Hydrocolloids*, 70, 345–355. 10.1016/j.foodhyd.2017.04.014.
- Srinivasa, P. C., Ramesh, M. N., Kumar, K. R., & Tharanathan, R. N. (2004). Properties of chitosan films prepared under different drying conditions. *Journal of Food Engineering*, 63(1), 79–85. 10.1016/S0260-8774(03)00285-1.
- Suhag, R., Kumar, N., Petkoska, A. T., & Upadhyay, A. (2020). Film formation and deposition methods of edible coating on food products: A review. *Food Research International*, 136, Article 109582 October. 10.1016/j.foodres.2020.109582.
- Sun, Q., Sun, C., & Xiong, L. (2013). Mechanical, barrier and morphological properties of pea starch and peanut protein isolate blend films. *Carbohydrate Polymers*, 98(1), 630–637. 10.1016/j.carbpol.2013.06.040.
- Tapia-Blácido, D. R., Sobral, P. J. d. A., & Menegalli, F. C. (2013). Effect of drying conditions and plasticizer type on some physical and mechanical properties of amaranth flour films. *LWT - Food Science and Technology*, 50(2), 392–400. 10.1016/j.lwt.2012.09.008.
- Tavares, L., Souza, H. K. S., Gonçalves, M. P., & Rocha, C. M. R. (2021). Physicochemical and microstructural properties of composite edible film obtained by complex coacervation between chitosan and whey protein isolate. *Food Hydrocolloids*, 113 June 2020. 10.1016/j.foodhyd.2020.106471.
- Tavassoli-Kafrani, E., Shekarchizadeh, H., & Masoudpour-Behabadi, M. (2016a). Development of edible films and coatings from alginates and carrageenans. *Carbohydrate Polymers*, 137, 360–374. 10.1016/j.carbpol.2015.10.074.
- Tomás, S. A., Bosquez-Molina, E., Stolik, S., & Sánchez, F. (2005). Effects of mesquite gumcandelilla wax based edible coatings on the quality of guava fruit (Psidium guajava L.). In Journal de Physique IV (Proceedings), 125, 889–892 EDP sciences.
- Tsai, M. J., & Weng, Y. M. (2019). Novel edible composite films fabricated with whey protein isolate and zein: Preparation and physicochemical property evaluation. *LWT*, 101, 567–574. 10.1016/j.lwt.2018.11.068.
- Uranga, J., Etxabide, A., Guerrero, P., & de la Caba, K. (2018). Development of active fish gelatin films with anthocyanins by compression molding. *Food Hydrocolloids*, 84, 313–320. 10.1016/j.foodhyd.2018.06.018.
- V, A. K., Srivastav, P. P., Pravitha, M., Hasan, M., Mangaraj, S., V, P., & Verma, D. K (2022). Comparative study on the optimization and characterization of soybean aqueous extract-based composite film using response surface methodology (RSM) and artificial neural network (ANN). *Food Packaging and Shelf Life*, 31, Article 100778. 10.1016/j.fpsl.2021.100778.
- V, A. K., Pravitha, M., Srivastav, P. P., Mangaraj, S., Pandiselvam, R., & Hasan, M (2021). Development of soy-based nanocomposite film: Modeling for barrier and mechanical properties and its application as cheese slice separator. *Journal of Texture Studies*. 10.1111/jtxs.12636.
- Valdés, A., Garcia-Serna, E., Martínez-Abad, A., Vilaplana, F., Jimenez, A., & Garrigós, M. C. (2020). Gelatin-Based Antimicrobial Films Incorporating Pomegranate (*Punica granatum L.*) Seed Juice by-Product. *Molecules*, 25(1), 166. 10.3390/molecules25010166.
- Valencia-Chamorro, S. A., Pérez-Gago, M. B., del Río, M. Á., & Palou, L. (2009). Effect of antifungal hydroxypropyl methylcellulose (HPMC)-lipid edible composite coatings on postharvest decay development and quality attributes of coldstored "Valencia" oranges. *Postharvest Biology and Technology*, 54(2), 72–79. 10.1016/j.postharvbio.2009.06.001.
- Valencia-Sullca, C., Vargas, M., Atarés, L., & Chiralt, A. (2018). Thermoplastic cassava starch-chitosan bilayer films containing essential oils. *Food Hydrocolloids*, 75, 107– 115. 10.1016/j.foodhyd.2017.09.008.
- Vedove, T. M. A. P. D., Maniglia, B. C., & Tadini, C. C. (2021). Production of sustainable smart packaging based on cassava starch and anthocyanin by an extrusion process. *Journal of Food Engineering*, 289, Article 110274. 10.1016/j.jfoodeng.2020.110274.
- Vlachogianni, T., & Valavanidis, A. (2014). Nanomaterials: Environmental pollution, ecolological risks and adverse health effects. Nano Sci. Nano Technol., 8(6), 208–226.
- Volpe, M. G., Coccia, E., Siano, F., Di Stasio, M., & Paolucci, M. (2019). Rapid evaluation methods for quality of trout (*Oncorhynchus mykiss*) fresh fillet preserved in an active edible coating. *Foods*, 8(4), 113. 10.3390/foods8040113.
- Wagh, Y. R., Pushpadass, H. A., Emerald, F. M. E., & Nath, B. S. (2014). Preparation and characterization of milk protein films and their application for packaging of Cheddar cheese. *Journal of Food Science and Technology*, 51(12), 3767–3775. 10.1007/s13197-012-0916-4.
- Wang, Kai, Wu, K., Xiao, M., Kuang, Y., Corke, H., Ni, X., & Jiang, F. (2017). Structural characterization and properties of konjac glucomannan and zein

blend films. International Journal of Biological Macromolecules, 105, 1096-1104. 10.1016/j.ijbiomac.2017.07.127.

- Wang, Kun, Wang, W., Ye, R., Liu, A., Xiao, J., Liu, Y., & Zhao, Y (2017). Mechanical properties and solubility in water of corn starch-collagen composite films: Effect of starch type and concentrations. *Food Chemistry*, 216, 209–216. 10.1016/j.foodchem.2016.08.048.
- Wang, L., Ding, J., Fang, Y., Pan, X., Fan, F., Li, P., & Hu, Q. (2020). Effect of ultrasonic power on properties of edible composite films based on rice protein hydrolysates and chitosan. Ultrasonics Sonochemistry, 65, Article 105049 February. 10.1016/j.ultsonch.2020.105049.
- Wang, Q., Lei, J., Ma, J., Yuan, G., & Sun, H. (2018). Effect of chitosan-carvacrol coating on the quality of Pacific white shrimp during iced storage as affected by caprylic acid. *International journal of biological macromolecules*, 106, 123–129. 10.1016/j.ijbiomac.2017.07.180.
- Wang, Yuemeng, Liu, A., Ye, R., Wang, W., & Li, X. (2015). Transglutaminase-induced crosslinking of gelatin-calcium carbonate composite films. *Food Chemistry*, 166, 414– 422. 10.1016/j.foodchem.2014.06.062.
- Wittaya, T. (2012). Protein-Based Edible Films: Characteristics and Improvement of Properties. In Ayman Amer Eissa (Ed.), Structure and Function of Food Engineering. InTech. 10.5772/48167.
- Xia, C., Wang, L., Dong, Y., Zhang, S., Shi, S. Q., Cai, L., & Li, J. (2015). Soy protein isolatebased films cross-linked by epoxidized soybean oil. RSC Advances, 5(101), 82765– 82771. 10.1039/c5ra15590h.
- Xu, X., Liu, H., Duan, S., Liu, X., Zhang, K., & Tu, J. (2019). A novel pumpkin seeds proteinpea starch edible film: Mechanical, moisture distribution, surface hydrophobicity, UVbarrier properties and potential application. *Materials Research Express*, 6(12), Article 125355, 10.1088/2053-1591/ab63f7.
- Xue, F., Gu, Y., Wang, Y., Li, C., & Adhikari, B. (2019). Encapsulation of essential oil in emulsion based edible films prepared by soy protein isolate-gum acacia conjugates. *Food Hydrocolloids*, 96, 178–189 May. 10.1016/j.foodhyd.2019.05.014.
- Yang, S., Wang, J., Wang, Y., & Luo, Y. (2016). Key role of collagen fibers orientation in casing-meat adhesion. *Food Research International*, 89, 439–447. 10.1016/j.foodres.2016.08.035.
- Yekta, R., Mirmoghtadaie, L., Hosseini, H., Norouzbeigi, S., Hosseini, S. M., & Shojaee-Aliabadi, S. (2020). Development and characterization of a novel edible film based on *Althaea rosea* flower gum: Investigating the reinforcing effects of bacterial nanocrystalline cellulose. *International Journal of Biological Macromolecules*, 158, 327–337. 10.1016/j.ijbiomac.2020.04.021.
- Yerramathi, B. B., Kola, M., Muniraj, B. A., Aluru, R., Thirumanyam, M., & Zyryanov, G. V. (2021). Structural studies and bioactivity of sodium alginate edible films fabricated through ferulic acid crosslinking mechanism. *Journal of Food Engineering*, 301, Article 110566.
- Yin, Y. C., Yin, S. W., Yang, X. Q., Tang, C. H., Wen, S. H., Chen, Z., Xiao, B. jie, & Wu, L. Y. (2014). Surface modification of sodium caseinate films by zein coatings. *Food Hydrocolloids*, 36, 1–8. 10.1016/j.foodhyd.2013.08.027.
- Yuan, G., Zhang, X., Tang, W., & Sun, H. (2015). CyTA-Journal of Food Effect of chitosan coating combined with green tea extract on the melanosis and quality of Pacific white shrimp during storage in ice. CyTA-Journal of Food, 14(1), 35–40. 10.1080/19476337.2015.1040459.
- Zhang, S., & Zhao, H. (2017). Preparation and properties of zein-rutin composite nanoparticle/corn starch films. *Carbohydrate Polymers*, 169, 385–392. 10.1016/j.carbpol.2017.04.044.
- Zhang, X., Xiao, G., Wang, Y., Zhao, Y., Su, H., & Tan, T. (2017). Preparation of chitosan-TiO2 composite film with efficient antimicrobial activities under visible light for food packaging applications. *Carbohydrate Polymers*, 169, 101–107. 10.1016/j.carbpol.2017.03.073.
- Zhang, T., Yu, Z., Ma, Y., Chiou, B.-S., Liu, F., & Zhong, F. (2022). Modulating physicochemical properties of collagen films by cross-linking with glutaraldehyde at varied pH values. *Food Hydrocolloids*, 124, Article 107270. 10.1016/j.foodhyd.2021.107270.
- Zhou, W., He, Y., Liu, F., Liao, L., Huang, X., Li, R., Zou, Y., Zhou, L., Zou, L., Liu, Y., Ruan, R., & Li, J. (2021). Carboxymethyl chitosan-pullulan edible films enriched with galangal essential oil: Characterization and application in mango preservation. *Carbohydrate Polymers*, 256, Article 117579. 10.1016/j.carbpol.2020.117579.
- Zoghi, A., Khosravi-Darani, K., & Mohammadi, R. (2020). Application of edible films containing probiotics in food products. *Journal of Consumer Protection and Food Safety*, 15(4), 307–320. 10.1007/s00003-020-01286-x.
- Zubeldía, F., Ansorena, M. R., & Marcovich, N. E. (2015). Wheat gluten films obtained by compression molding. *Polymer Testing*, 43, 68–77. 10.1016/j.polymertesting.2015.02.001.
- Zuo, G., Song, X., Chen, F., & Shen, Z. (2019). Physical and structural characterization of edible bilayer films made with zein and corn-wheat starch. *Journal of the Saudi Society* of Agricultural Sciences, 18(3), 324–331. 10.1016/j.jssas.2017.09.005.
- USFDA. Part 172 Food additives permitted for direct addition to food for human consumption. CFR - Code of Federal Regulations Title 21. 2014. (Accessed 15 March 2022).